

NEMATOLOGICAL REVIEWS—RESENAS NEMATOLOGICAS  
 USE OF THE ENTOMOGENOUS NEMATODE, *NEOAPLECTANA  
 CARPOCAPSAE* WEISER (STEINERNEMATIDAE: RHABDI-  
 TIDA), IN LATIN AMERICA

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ABSTRACT

Wassink, H., and G.O. Poinar, Jr. 1984. Use of the entomogenous nematode, *Neoaplectana carpocapsae* Weiser (Steinernematidae: Rhabditida), in Latin America. *Nematropica* 14:97-109.

The present report was prepared to collect and analyze the literature dealing with the testing of *Neoaplectana carpocapsae* against insect pests of Latin America. A total of 97 species of Latin American insects from 11 orders were shown to be susceptible to *N. carpocapsae*. Native strains of *N. carpocapsae* occur in Latin America and these and isolates from other parts of the world should be tested against economically important insect pests in Latin America.

The infective stages of *N. carpocapsae* occur in the soil and will be most effective against soil-inhabiting insects. Since rapid desiccation will destroy the infective stages in a short period, application of *N. carpocapsae* to exposed surfaces should be done with an additive to retard water loss. Field trials show that *N. carpocapsae* has real potential for use as a biological control agent in pest management programs.

*Additional key words:* biological control, pest management.

RESUMEN

Wassink, Han, y George O. Poinar, Jr. 1984. Uso del nematodo entomógeno, *Neoaplectana carpocapsae* Wesier (Steinernematidae: Rhabditida), en la America Latina. *Nematropica* 14:97-109.

Este trabajo consistió en una recopilación y análisis de la literatura relacionada con el uso de *Neoaplectana carpocapsae* contra plagas insectiles de la America Latina. Un total de 97 especies de insectos que comprenden 11 órdenes resultaron ser susceptibles al *N. carpocapsae*. Las cepas naturales de *N. carpocapsae* que ocurren en la America Latina, junto con aislamientos de otras partes del mundo deban ser evaluades contra plagas de importancia economica de la America Latina.

Los estadios infectivos de *N. carpocapsae* ocurren en el suelo, por lo que su uso resulta más efectivo contra insectos que habitan en la suelo. Debido a que los estadios infectivos son destruidos rapidamente por la desecación, las aplicaciones de *N. carpo-*

*capsae* en áreas no protogidas deben ser efectuadas usando algun, aditivo que retarde al pérdida de la humedad. Ensayos de campo indican que *N. carpocapsae* tiene un potencial efectivo para ser usado como un agente biológico en los programas de manejo de plagas.

*Palabras claves adicionales:* control biológico, manejo de plagas.

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## INTRODUCTION

Neoaplectanid nematodes constitute one of the most important biological control agents for soil insects (11,16,17,27,29,30,31,37). The most widely used species is *Neoaplectana carpocapsae* Weiser, which is composed of many geographical strains (30). The infective third-stage juveniles of *N. carpocapsae* enter their hosts via natural body openings (mouth, spiracles, anus) and then penetrate into the hemocoel. The infective stages carry a mutualistic bacterium, *Xenorhabdus nematophilus* (36), in the lumen of their intestine and release this bacterium into the host's body cavity. The bacteria multiply and kill the host within 48 hr. The nematodes then develop in the bacterial-ridden insect remains, undergo one or two generations, and form infective stages that return to the environment.

*Neoaplectana carpocapsae* has a wide host range (22,30) and has been tested against medically important insects (mosquitoes, blackflies, tsetse flies, triatomid bugs) as well as agriculturally important soil pests.

Use of *N. carpocapsae* in Latin America has been restricted mainly to the laboratory with a few field trials. These are listed in Table 1. A few of these tests were conducted in countries outside Latin America, but since they included insects from this region, they are included here. In some cases, information on the results of the tests and stages used was not reported.

## POTENTIAL OF *N. CARPOCAPSAE* AGAINST INSECT PESTS IN LATIN AMERICA

*Medically important insects.* Although working outside Latin America, both Skierska and Szadziewska (34) and Minter and Oswald (24) showed that both *Rhodnius prolixus* and *Triatoma infestans*, vectors of American trypanosomiasis, were susceptible to infection by *N. carpocapsae*. These results were also supported by studies in Mexico (38), where the local species were also found to be susceptible.

*Agriculturally important insects.* The cutworm pest of corn, *Spodoptera frugiperda*, is especially susceptible to *N. carpocapsae* in the laboratory (22). When used in the field in Colombia at the rate of 4000 nematodes per corn plant, 50-60% of the larvae were destroyed (21).

Table 1. Insects used as hosts for *Neoplectana carpocapsae* in laboratory and field trials in Latin America.

Insect	Trials		Country	Results#	Reference
	Stage <sup>ø</sup>	(lab./field)			
ODONATA					
Aeschnidae					
<i>Anax junius</i>	A	lab.	Guadeloupe	++++	22
PHASMIDA					
Phasmidae					
<i>Pseudobacteria crudelis</i>	A	lab.	Guadeloupe	—	22
DICTYOPTERA					
Blatellidae					
<i>Blatella germanica</i>	A	lab.	Guadeloupe	—	22
<i>Blatella germanica</i>	A	lab.	Poland	94.77% mort.	34
ISOPTERA					
Termitidae					
<i>Nasutitermes costalis</i>	A	lab.	Guadeloupe	100% mort. in 3 days	22
ORTHOPTERA					
Gryllidae					
<i>Gryllus assimilis</i>	A	lab.	Guadeloupe	++++	22
Nemobiidae					
<i>Hygronemobius</i> sp.	A	id.	id.	++++	id.
Gryllotalpidae					
<i>Neocurtilla hexadactyla</i>	A	id.	id.	+++	id.
Phaneropteridae					
<i>Turpilia rugulosa</i>	A	id.	id.	++++	id.
HEMIPTERA					
HETEROPTERA					
Belostomatidae					
<i>Belostoma boscii</i>	A	lab.	Guadeloupe	—	22
Coreidae					
<i>Phthia picta</i>	L	lab.	Guadeloupe	++++	22
Corixiidae <sup>z</sup>	A	id.	id.	—	id.
Notonectidae					
<i>Buenna antigone</i>	A	id.	id.	—	id.
Pentatomidae					
<i>Nezara viridula</i>	A	id.	id.	++++	id.
Pyrrhocoridae					
<i>Pyrrhocorus</i> sp.	A	lab.	id.	++++	id.
<i>Dysdercus cingulatus</i>	A	lab.	id.	+	id.
<i>Dysdercus peruvianus</i>	A	field	Peru	22-36% mort.	35
<i>Dysdercus peruvianus</i>	A	lab.	id.	100% mort. in 3 days	35

Table 1. Insects used as hosts for *Neoapectana carpocapsae* in laboratory and field trials in Latin America (continued).

Insect	Trials		Country	Results#	Reference
	Stage <sup>o</sup>	(lab./field)			
<b>Reduviidae</b>					
<i>Rhodnius prolixus</i>	A	lab.	Poland	75-100% mort.	34
<i>Rhodnius prolixus</i>	A	lab.	Great Britain	20-100% mort.	24
<i>Rhodnius prolixus</i>	A	field	Venezuela	nematodes can effectively kill the bugs in the field	25
	N,A	lab.	Mexico	+++	38
<i>Triatoma infestans</i>	N,A,L	lab.	Poland	95% mort. in 3 days	34
<i>Triatoma barberi</i>	N,A	lab.	Mexico	++	38
<i>Triatoma mazzotti</i>	N,A	lab.	Mexico	++	38
<b>Tingidae</b>					
<i>Corythaica cyathiolis</i>	A	lab.	Guadeloupe	++++	22
<b>HEMIPTERA</b>					
<b>HOMOPTERA</b>					
<b>Aphididae</b>					
<i>Brevicoryne brassicae</i>	A	lab.	Guadeloupe	+	22
<i>Cicadidae</i> <sup>o</sup>	A	id.	id.	—	id.
<b>Margarodidae</b>					
<i>Margarodes vitium</i>	L	id.	Chile	++	9
<b>Pseudococcidae</b>					
<i>Ferrisia</i> sp.	A	id.	id.	+	id.
<i>Pseudococcus</i> sp.	A	id.	id.	+	id.
<b>NEUROPTERA</b>					
<b>Chrysopidae</b>					
<i>Chrysopa collaris</i>	A	lab.	Guadeloupe	++++	22
<b>Myrmeleonidae</b>					
<i>Myrmeleon</i> sp.	A	lab.	id.	—	id.
<b>COLEOPTERA</b>					
<b>Bostrychidae</b>					
<i>Dinoderus minutus</i>	A	lab.	Guadeloupe	74% mort. in 3 days	22
<i>Dinoderus minutus</i>	A	lab.	id.	++++	id.
<b>Cerambycidae</b>					
<i>Eburia octomaculata</i>	A	lab.	Guadeloupe	++++	id.
<i>Chlorida festiva</i>	A	id.	id.	++++	id.
<b>Chrysomelidae</b>					
<i>Diabrotica balteata</i>	L	lab.	Guadeloupe	100% mort. in 6-7 days	id.
<b>Coccinellidae</b>					
<i>Coleomegylla</i> sp.	A	lab.	Guadeloupe	++++	id.

Table 1. Insects used as hosts for *Neoaeplectana carpocapsae* in laboratory and field trials in Latin America (continued).

Insect	Trials		Country	Results <sup>y</sup>	Reference
	Stage <sup>x</sup>	(lab./field)			
<i>Cycloneda sanguinea</i>	A	lab.	id.	+++	id.
Curculionidae					
<i>Anthonomus vestitus</i>	A	lab.	Peru	100% mort. in 3 days	35
<i>Lophotus phaleratus</i>	L	lab.	Chile	+++	9
<i>Cosmopolites sordidus</i>	A	id.	Guadeloupe	++++	22
<i>Diaprepes abbreviatus</i>	A	id.	id.	+++	id.
<i>Diaprepes famelicus</i>	A	id.	id.	++++	id.
<i>Diaprepes marginatus</i>	A	id.	id.	++++	id.
<i>Heilipus latro</i>	L	id.	id.	++++	id.
<i>Metamasius hemipterus</i>	A	id.	id.	++++	id.
<i>Pantomorus</i> sp.	L	id.	Chile	+++	9
Cleridae					
<i>Madoniella pici</i>	A	lab.	Guadeloupe	++++	22
Dysticidae					
<i>Cybister</i> sp.	A	lab.	Guadeloupe	—	id.
Elateridae					
<i>Chalcolepidius obscurus</i>	A	lab.	Guadeloupe	+++	id.
<i>Pyrophorus</i>					
<i>phosphorescens</i>	A	lab.	id.	—	id.
Hydrophilidae					
<i>Hydrophilus</i> sp.	A	lab.	id.	—	id.
Passalidae					
<i>Passalus unicornis</i>	A	lab.	Guadeloupe	++++	id.
<i>Paxillus puniticollis</i>	A	id.	id.	+++	id.
Scarabaeidae					
<i>Anomala insularis</i>	A	lab.	Guadeloupe	—	22
<i>Cyclocephala</i> sp.	L	lab.	id.	+	id.
<i>Dynastes hercules</i>	A	lab.	id.	—	id.
<i>Epilachna varivestis</i>	A	lab.	Mexico		4
<i>Hylamorpha elegans</i>	L	lab.	Chile	+++	9
<i>Ligyris cuniculus</i>	A	lab.	Guadeloupe	—	22
<i>Macraspis tristis</i>	A	lab.	id.	++++	id.
<i>Phyllophaga</i>					
<i>patrueloides</i>	A	lab.	id.	+	id.
<i>Phyllophaga pleci</i>	L	lab.	id.	—	id.
<i>Phyllophaga</i> sp.	L	lab.	Mexico	++	4
<i>Premnotrypes vorax</i>	L	lab.	Colombia	+++	3
Scolytidae					
<i>Hexacolus guyanensis</i>	L	lab.	Guadeloupe	++++	22
<i>Hexacolus guyanensis</i>	P	id.	id.	++++	id.
<i>Hexacolus guyanensis</i>	A	id.	id.	++++	id.
<i>Dendroctonus adjunctus</i>	A	lab.	Mexico		4

Table 1. Insects used as hosts for *Neoaplectana carpocapsae* in laboratory and field trials in Latin America (continued).

Insect	Trials		Country	Results <sup>#</sup>	Reference
	Stage <sup>#</sup>	(lab./field)			
<b>Tenebrionidae</b>					
<i>Zophobas atratus</i>	A	lab.	Guadeloupe	++++	22
<b>DIPTERA</b>					
<b>Chironomidae<sup>2</sup></b>					
	L	lab.	Guadeloupe	100% m. in 3 days	22
<b>Calliphoridae</b>					
<i>Ornidia obesa</i>	A	lab.	Guadeloupe	--	id.
<b>Culicidae</b>					
<i>Aedes aegypti</i>	L	lab.	Guadeloupe	100% m. in 3 days	id.
<b>Drosophilidae</b>					
<i>Drosophila repleta</i>	L	lab.	Guadeloupe	27% m. in 3 days	id.
<b>Muscidae</b>					
<i>Musca domestica</i>	E	lab.	Guadeloupe	--	id.
<i>Musca domestica</i>	L	id.	id.	--	id.
<i>Musca domestica</i>	A	id.	id.	+++	id.
<b>Tachinidae</b>					
<i>Metagonistylum minense</i>	A	lab.	Guadeloupe	++++	id.
<b>Tipulidae</b>					
<i>Tipula</i> sp.	L	lab.	Chile	--	9
<b>Trypetidae</b>					
<i>Anastrepha ludens</i>	A	lab.	Mexico	+	4
<b>LEPIDOPTERA</b>					
<b>Bombycidae</b>					
<i>Bombyx mori</i>	L	lab.	Peru	susceptible	35
<b>Gelechiidae</b>					
<i>Phthorimaea opercutella</i> ( <i>Gnorimoschema</i> )	L	lab.	Chile	++++	9
<i>Scrobipalpula absoluta</i>	L	field	Colombia	after 5-10 days good control	33
<b>Geometridae</b>					
<i>Oxydia trychiata</i>	L,P	lab.	Colombia	all stages were killed	8
<b>Hepialidae</b>					
<i>Maculella noctuides</i>	L	lab.	Chile	+++	9
<b>Hesperiidae</b>					
<i>Urbanus proteus</i>	L	lab.	Guadeloupe	70% in 7 days	22
<i>Xanthopastis timais</i>	L	lab.	id.	--	id.
<b>Hyponomeutidae</b>					
<i>Plutella maculipennis</i>	L	lab.	Guadeloupe	10% m. in 6-7 days	id.

Table 1. Insects used as hosts for *Neoaplectana carpocapsae* in laboratory and field trials in Latin America (continued).

Insect	Trials		Country	Results <sup>#</sup>	Reference
	Stage <sup>w</sup>	(lab./field)			
<b>Noctuidae</b>					
<i>Agrotis</i> sp.	L	lab.	Chile	+++	9
<i>Gonodonta</i> sp.	A	lab.	Guadeloupe	—	22
<i>Heliothis virescens</i>	L	lab.	Peru	more susceptible	35
<i>Mocis punctularis</i>	L	lab.	Guadeloupe	++++	22
<i>Prodenia eridania</i>	L	lab.	id.	++++	id.
<i>Scotia subterranea</i>	L	lab.	id.	100% in 6-7 days	id.
<i>Spodoptera frugiperda</i>	L	lab.	id.	94% in 4-5 days	id.
<i>Spodoptera frugiperda</i>	L	field	Colombia	50-60% decrease	21
<i>Spodoptera frugiperda</i>	L	field	Colombia	35% parasitism	7
<b>Oleuthreutidae</b>					
<i>Epimotia aporema</i>	L	lab.	Chile	++++	22
<b>Pyralidae</b>					
<i>Achroia grisella</i>	L	lab.	Peru	100% m. in 1 day	35
<i>Diaphania hyalinata</i>	L	lab.	Guadeloupe	100% m. in 3 days	22
<i>Diatraea saccharalis</i>	L	lab.	id.	96% m. in 6-7 days	id.
<i>Elasmopalpus lignosellus</i>	L	lab.	id.	++	id.
<i>Ephestia kuehniella</i>	L	lab.	id.	++++	id.
<i>Galleria mellonella</i>	L	lab.	id.	++++	id.
<i>Hypsipyla grandella</i>	L	lab.	id.	++++	id.
<i>Mescinia peruella</i>	L	lab.	Peru	100% in 2 days	35
<i>Pococera atramentalis</i>	L	lab.	id.	susceptible	id.
<i>Zinkenja fascialis</i>	L	lab.	Guadeloupe	27% m. in 6-7 days	id.
<b>Pyraustidae</b>					
<i>Mesocondyla concordalis</i>	L	lab.	Guadeloupe	++++	id.
<b>Sphingidae</b>					
<i>Pseudosphinx tetrio</i>	L	lab.	Guadeloupe	++++	id.
<b>Zeuzeridae</b>					
<i>Langsdorfia</i> sp.	L	lab. & field	Chile	++++	9
<b>HYMENOPTERA</b>					
<b>Diprionidae</b>					
<i>Zadiprion vallicola</i>	L	lab.	Mexico	++	4
<b>Formicidae</b>					
<i>Acromyrmex</i>					
<i>octospinosus</i>	L	lab.	Guadeloupe	++++	22
<i>Acromyrmex</i>					
<i>octospinosus</i>	N	lab.	id.	+	id.
<i>Acromyrmex</i>					
<i>octospinosus</i>	A	lab.	id.	—	id.

Table 1. Insects used as hosts for *Neoaplectana carpocapsae* in laboratory and field trials in Latin America (continued).

Insect	Stage <sup>w</sup>	Trials		Results <sup>y</sup>	Reference
		(lab./field)	Country		
<i>Acromyrmex octospinosus</i>	A,L	lab. & field	id.	inactivation of the nests wild and art.	20
<i>Camponotus</i> sp.	A	lab.	Guadeloupe	+++	22
<i>Camponotus</i> sp.	L	lab.	id.	+++	id.
<i>Camponotus</i> sp.	A	lab.	id.	82% m. in 6-7d.	id.
<i>Camponotus</i> sp.	A	lab.	id.	30% m. in 6-7d.	id.
<i>Solenopsis geminata</i>	A	lab.	id.	100% m. in 3 d.	id.
<i>Solenopsis richteri</i>	A,L	field	U.S.A.	80% inactivation of the nids in 90 days	19
<i>Solenopsis invicta</i>	A,L	field	U.S.A.	80% inactivation of the nids in 90 days	id.
Scoliidae					
<i>Campomeris dorsata</i>	A	lab.	Guadeloupe	+++	22
Sphegidae					
<i>Sphex caliginosus</i>	A	lab.	Guadeloupe	+++	id.
Vespidae					
<i>Polistes</i> sp.	L	lab.	Guadeloupe	+++	id.
Xylocopidae					
<i>Xylocopa mordax</i>	A	lab.	Guadeloupe	+++	id.

<sup>w</sup>A = adult, L = larva, E = egg, P = pupa, N = juvenile.

<sup>y</sup>(-) = no infection, (+) = very light infection, (++) = light infection, (+++) = susceptible, (++++) = very susceptible.

<sup>z</sup>Not identified.

Laboratory tests simply show whether the host is susceptible under ideal conditions. Other factors, especially the problem of desiccation of inoculum, should be taken into account when conducting field trials. This may be why the susceptible insects, *Scrobipalpula absoluta* (Lepidoptera, Gelechiidae) in Colombia and *Dysdercus peruvianus* (Heteroptera, Pyrrhocoridae) in Peru were not controlled in field trials (33,35).

At least two strains of *N. carpocapsae* have been recovered from plant pests in Latin America. One is the Mexican strain, originally recovered from larvae of *Laspeyresia pomonella* (codling moth) from Allende, Chihuahua, Mexico by L. Caltagirone (30). The second was found in Baliroche, Argentina from larvae of the alfalfa pests, *Graphognathus leucoloma*, *Naupactus devieus*, and *Pantomorus auripes* (Coleoptera, Curculionidae) (1,2).

*Other insect pests.* Ants are also susceptible to *N. carpocapsae* and



Kermarrec (20) showed that when applied to the nests of *Acromyrmex octospinosus*, the nematodes inactivated the hosts and were able to kill the larvae.

Adults of *Solenopsis geminata* were found to be susceptible to *N. carpocapsae* (22), and tests against the imported fire ant, *Solenopsis richteri*, and *S. invicta* in the United States gave interesting results. Using a dose of 1,000,000 nematodes per mound, it was possible to inactivate 80% of the mounds (19). The termite, *Nasutitermis costalis*, was very susceptible in laboratory tests (22). The cypress pest, *Oxydia trychiata* (Lepidoptera, Geometridae), was found to be very susceptible to *N. carpocapsae* under simulated field conditions in Colombia with nematode reproduction occurring in all stages (8).

### PROPAGATION OF *N. CARPOCAPSAE*

The easiest method of rearing *N. carpocapsae* and other neoaplectanid nematodes is with the use of live insects. In most laboratories, the larvae of the wax moth, *Galleria mellonella*, is used. The nematodes can also be grown on artificial media such as dog food agar (18) or animal protein on polyeder polyurethane sponges (5). Some researchers have used the adult stage of insects as hosts. Lindegren et al. (23) grew his material in adults of the naval orangeworm, *Amyelois transitella*.

*Improving field applications.* One of the problems with using nematodes in the field is their tendency to dry out and die. This is one reason why many experiments have failed (6). In addition, it has been shown that ultraviolet and natural sunlight can kill *Neoaplectana* (12,15). Certain adjuvants, such as paraamino-benzoic acid, can be used to protect the nematodes against light when used on exposed surfaces (14). To avoid both the problems of desiccation and UV light, it is advisable to make applications in the evening when the humidity tends to be highest.

### FUTURE PROSPECTS

There are many possibilities for the use of neoaplectanid nematodes in Latin America. Many soil pests on potato, corn, alfalfa, and other crops are susceptible to nematodes, thus avoiding many problems associated with the use of chemical insecticides. An attempt should be made to discover new Latin American strains of *Neoaplectana* (and other related genera such as *Heterorhabditis*). This can be done by removing soil and placing it in containers in the laboratory containing susceptible insects. When the insects are killed by nematodes in the soil, they can be removed and held until the infective stages emerge.

Some potential above-ground pests that might be controlled with

neoplectanid nematodes are the cocoa canker beetle, *Xyleborus ferrugineus* (Coleoptera, Scolytidae), and the oil palm caterpillar, *Castnia daedalus* (Lepidoptera, Castniidae).

*Neoplectana carpocapsae* has been exempted from registration by the Environmental Protection Agency in the United States since tests have shown that the nematodes cannot infect mammals and birds (13,32).

In summary, the following steps should be undertaken to progress with this means of biological control:

1. A search should be conducted for native species of *Neoplectana*.
2. Tests should be made with soil insects and those burrowing inside plants since these are likely candidates for field trials.
3. Anti-evaporation retardants should be used when applying nematodes against foliage feeding insects.
4. Consider the use of nematodes as one factor in an integrated control program, especially when soil insects are involved.

#### LITERATURE CITED

1. AHMAD, R. 1974. Investigations on the white-fringed weevils *Nau-pactus durius* Boh. and *Pantomorus auripes* Hustache (Coleoptera, Curculionidae) and their natural enemies in Argentina. Tech. Bull. CIBC 17:37-51.
2. AHMAD, R. 1974. Studies on *Graphognathus leucoloma* Boh. (Coleoptera, Curculionidae) and its natural enemies in the central provinces of Argentina. Tech. Bull. CIBC 17:19-28.
3. AMAYA, L.M., and E. BUSTAMANTE. 1975. Control microbiológico de tres coleópteros plagas del suelo en Colombia. Revista (Colombia) 10:269.
4. ALATORRE-ROSAS, R. 1971. El uso de *Neoplectana dutkyi* Jackson, 1965 (DD-136), Nematoda, Neoplectanidae en la lucha biológica contra algunas plagas de insectos. Thesis, Universidad Nacional Autónoma de Mexico, Mexico City, 61 pp.
5. BEDDING, R.A. 1981. Low cost *in vitro* mass production of *Neoplectana* and *Heterorhabditis* species (Nematoda) for field control of insect pests. Nematologica 27:109-114.
6. BENHAM, G. S., and G. O. POINAR, Jr. 1973. Tabulation and evaluation of recent field experiments using the DD-136 strain of *Neoplectana carpocapsae* Weiser: A review. Exp. Parasitol. 33:248-252.
7. BENJUMEA, M.H., H. GIRALDO C., and O. CASTANO P. 1978. Evaluación de métodos de cultivo para el nematodo *Neoplectana carpocapsae* y el ensayo de campo para el control de *Spodoptera frugiperda* en maíz. P. 26 in Resúmenes de los trabajos presentados al V

- Congreso de la Sociedad Colombiana de Entomología, 26-28 July, 1978. Ibaqué, Colombia.
8. BUSTILLO, A.E. 1976. Patogenicidad del nematodo *Neoaplectana carpocapsae* en larvas, prepupas y pupas de *Axydia trychiata*. Rev. Col. de Entomol. 2:139-144.
  9. DUTKY, S.R., J.V. THOMPSON, and W.S. HOUGH. 1962. A new nematode parasite of codling moth showing promise in insect control. Susceptibility of various insects to DD-136 nematode. 8th Int. Congr. Microbiol. Montreal, Canada (Mimeo. Abstr.) 6 pp. (with additional comments by L. Caltagirone).
  10. EL KADI M., K. 1977. Contribuicao ao dos nematoides entomogenos ao controle de insetos. Soc. Brasil Nemat. Public. No. 2:75-80.
  11. FINNEY, J.R. 1981. Potential of nematodes for pest control. Pp. 603-620 in H.D. Burgess (ed.), Microbial control of pests and plant diseases 1970-1980. Academic Press, London and New York, 949 pp.
  12. GAUGLER, R., and G.M. BOUSCH. 1978. Effects of ultraviolet radiation and sunlight on the entomogenous nematode *Neoaplectana carpocapsae*. J. Invertebr. Path. 32:291-296.
  13. GAUGLER, R., and G.M. BOUSCH. 1979. Nonsusceptibility of rats to the entomogenous nematode *Neoplectana carpocapsae*. Environ. Entomol. 8:658-660.
  14. GAUGLER, R., and G.M. BOUSCH. 1979. Laboratory tests on ultraviolet protectants of an entomogenous nematode. Environ. Entomol. 8:810-813.
  15. GAUGLER, R., and G.M. BOUSH. 1979. Effect of radiation on the entomogenous nematode *Neoaplectana carpocapsae*. J. Invertebr. Path. 33:121-123.
  16. GAUGLER, R. 1981. Biological control potential of Neoplectanid nematodes. J. Nematol. 13:241-249.
  17. GORDON, R., and J.M. WEBSTER. 1974. Biological control of insects by nematodes. Helminthological Abstracts, Series A, Animal and Human Helminthology 43:327-349.
  18. HARA, A.H., J.E. LINDEGREN, and H.K. KAYA. 1981. Monoxenic mass-production of the entomogenous nematode, *Neoaplectana carpocapsae* Weiser on dog food-agar medium. USDA/SEA, Adv. Agric. Technol., Western Ser. No. 16, Oakland, Calif.
  19. JOUVENAZ, D.P., S.C. LOFGREN, and W.A. BANKS. 1981. Biological control of imported fire ants: A review of current knowledge. Bull. Entomol. Soc. Am. 27:203-208.
  20. KERMARREC, A. 1975. Etude des relations synécologiques entre les nématodes et la fourmi manioc: *Acromyrmex octospinosus* Reich. Ann. Zool. Ecol. Anim. 7:27-44.

21. LANDSZABAL A., J., F. FERNANDEZ A., and A. FIGUERO P. 1973. Control biológico de *Spodoptera frugiperda* (J.E. Smith), con el nemátodo: *Neoaplectana carpocapsae* en maíz (*Zea mays*). Acta Agronómica (Palmira) 23:41-70.
22. LAUMOND, C., H. MAULEON, and A. KERMARREC. 1979. Données nouvelles sur le spectre d'hotes et le parasitisme du nématode entomophage *Neoaplectana carpocapsae*. Entomophaga 24:13-27.
23. LINDEGREN, J.E., D.F. HOFFMAN, S.S. COLLIER, and R.D. FRIES. 1979. Propagation and storage of *Neoaplectana carpocapsae* Weiser using *Amyelois transitella* (Walker) adults. USDA/SEA, Adv. Agric. Technol., Western Series No. 3, Berkeley, Calif.
24. MINTER, D.M., and W.J.C. OSWALD. 1980. The nematode *Neoaplectana* and its possible role in the biological control of triatomine bugs and other vectors. 10th Int. Congr. Trop. Med. Mal., Manila: abstract number 121:80.
25. OSWALD, W.J.C., and D.M. MINTER. 1980. The nematode *Neoaplectana carpocapsae*: as a potential biological control agent for triatomine bugs and other insects. Parasitology 80:43-44.
26. POINAR, G.O., Jr. 1971. Use of nematodes for biological control of insects. Pp. 181-201 in H.D. Burges and N.W. Hussey (eds.), Microbial control of insects and mites. Academic Press, London and New York. 861 pp.
27. POINAR, G.O., Jr. 1972. Nematodes as facultative parasites of insects. Ann. Rev. Entomol. 17:103-122.
28. POINAR, G.O., Jr. 1973. Entomogenous nematodes—the development of a neglected field. P. 86 in Abstracts of Papers Intern. Coll. Insect Path. and Microbial Control, September 1973, Oxford, England.
29. POINAR, G.O., Jr. 1975. Entomogenous nematodes—a manual and host list of insect-nematode associations. E.J. Brill, Leiden, The Netherlands. 317 pp.
30. POINAR, G.O., Jr. 1979. Nematodes for biological control of insects. CRC Press, Inc. Boca Raton, Florida. 277 pp.
31. POINAR, G.O., Jr. 1983. The natural history of nematodes. Prentice Hall, Englewood Cliffs, N.J. 323 pp.
32. POINAR, G.O., Jr., G.M. THOMAS, S.B. PRESSER, and J.L. HARDY. 1982. Inoculation of entomogenous nematodes, *Neoaplectana* and *Heterorhabditis* and their associated bacteria *Xenorhabdus* spp. into chicks and mice. Environ. Entomol. 11:137-138.
33. PRADA, R.M.A., and J. GUTIERREZ P. 1976. Contribucion preliminar al control microbiologico de *Scrobipalpula absoluta* (Meyrick) con *Neoaplectana carpocapsae* Weiser y *Bacillus thuringiensis* Berl. en tomate *Lycopersicum esculentum*, Mill. Acta Agronomica

- (Palmira) (1974) publ. 1976, 24:116-137.
34. SKIERSKA, B. and M. SZADZIEWSKA. 1976. Laboratory tests for the usability of the entomophilic nematodes. Steinernematidae, Chitwood and Chitwood 1937, in biological control of some noxious arthropods. Bull. Inst. Mar. Trop. Med. Gdynia 27:207-227.
  35. TANG, J.L. 1958. Notas generales sobre nematodos portadores de bacterias como un método de control biologico. Rev. Peru. Entomol. 1:19-22.
  36. THOMAS, G.M., and G.O. POINAR, Jr. 1979. *Xenorhabdus* gen. nov., a genus of entomopathogenic, nematophilic bacteria of the family Enterobacteriaceae. Intern. J. Syst. Bact. 29:352-360.
  37. WELCH, H.E. 1965. Entomophilic nematodes. Ann. Rev. Entomol. 10:275-302.
  38. ZARATE, L., and G.O. POINAR, Jr. 1984. Laboratory tests of *Neoalectana* and *Heterorhabditis* nematodes against populations of triatomid bugs in Mexico (en Preparation).

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