

## STUDIES ON SOME SPECIES OF THE *XIPHINEMA AMERICANUM* GROUP (NEMATODA, DORYLAIMIDA) OCCURRING IN FLORIDA

F. Lamberti, F. De Luca, S. Molinari, L.W. Duncan, A. Agostinelli, M.I. Coiro, D. Dunn and V. Radicci

*Istituto di Nematologia Agraria, C.N.R., 70126 Bari, Italy*  
*Citrus Research and Education Center, University of Florida Lake Alfred, Florida 33850, U.S.A.*

**Summary.** The study of the morphometrics of putative *Xiphinema americanum* group populations collected in various *habitats* in Florida, United States of America, revealed the presence of four species: *X. citricolum* Lamberti *et* Bleve-Zacheo, 1979; *X. floridae* Lamberti *et* Bleve-Zacheo, 1979; *X. georgianum* Lamberti *et* Bleve-Zacheo, 1979 and *X. laevistriatum* Lamberti *et* Bleve-Zacheo, 1979. The relationships of these species with *X. americanum sensu stricto* and similar species are discussed. It is shown for the first time that *X. citricolum* and *X. laevistriatum* have three juvenile stages. The SOD isozyme iso-electrofocusing patterns also separated the studied populations into four groups; however, it confirmed only partially the morphotypes. The analysis of the ribosomal DNA of seven populations characterized three groups identifying *X. floridae*, *X. citricolum* and *X. laevistriatum*. However, one population morphometrically identified as *X. laevistriatum* presented a pattern as *X. citricolum*.

*Xiphinema americanum sensu lato* is wide-spread in Florida (Tarjan, 1974) and several putative species of the group were originally described from the State (Lamberti and Bleve-Zacheo, 1979). They are *X. citricolum* Lamberti *et* Bleve-Zacheo, 1979; *X. floridae* Lamberti *et* Bleve-Zacheo, 1979; *X. intermedium* Lamberti *et* Bleve-Zacheo, 1979; *X. laevistriatum* Lamberti *et* Bleve-Zacheo, 1979 and *X. tarjanense* Lamberti *et* Bleve-Zacheo, 1979. Populations collected in Florida were also used, although not as type populations, in the original description of *X. diffusum* Lamberti *et* Bleve-Zacheo, 1979 and of *X. georgianum* Lamberti *et* Bleve-Zacheo, 1979. Later, also *X. luci* Lamberti *et* Bleve-Zacheo, 1979 and *X. sberi* Lamberti *et* Bleve-Zacheo, 1979 have been reported in Florida (Lamberti *et al.*, 2000). However, some identifications require confirmation (Robbins, 1993) and the validity of various species attributed to the *X. americanum* group is questioned (Luc *et al.*, 1998).

Several populations belonging to the *X. americanum* group were collected during a nematode survey carried out in Florida during October - November 1996. They were studied biometrically and, for some population, the superoxide dismutase (SOD) activity and the DNA of the ITS region were characterized. When available, the juvenile stages were determined and illustrated.

### MATERIALS AND METHODS

Soil samples were collected from the rhizosphere of cultivated plants and in natural *habitats*. Nematodes were extracted by a wet sieving technique. Specimens for biometric studies were fixed in 5% boiling formalin and mounted in anhydrous glycerin. Measurements were taken with the aid of a camera lucida.

SOD isozymes were separated by iso-electric-focusing on lots of 15-20 specimens (Molinari *et al.*, 1997).

Genomic DNA was amplified with the ITS primers (Molinari *et al.*, 1997) and the PCR amplification products were digested with the restriction enzymes *Alu* I, *Bam* HI, *Dde* I, *Hinf* I, *Rsa* I and *Xba* I (Lamberti *et al.*, 1999). DNA of four specimens from each population was analyzed to confirm the reproducibility of the fragment profiles.

### RESULTS AND DISCUSSION

The 13 populations studied (Table I) were identified as four species.

#### *XIPHINEMA CITRICOLUM* Lamberti *et* Bleve-Zacheo, 1979 (Tables I, II and VI; Figs 1-3, 9 and 10)

*Xiphinema citricolum* was the most commonly encountered species (Table I). Its morphometrics differ from the original description (Lamberti and Bleve-Zacheo, 1979) in its shorter odontostyle and thinner body profile. However, the old specimens of the original description were partially collapsed.

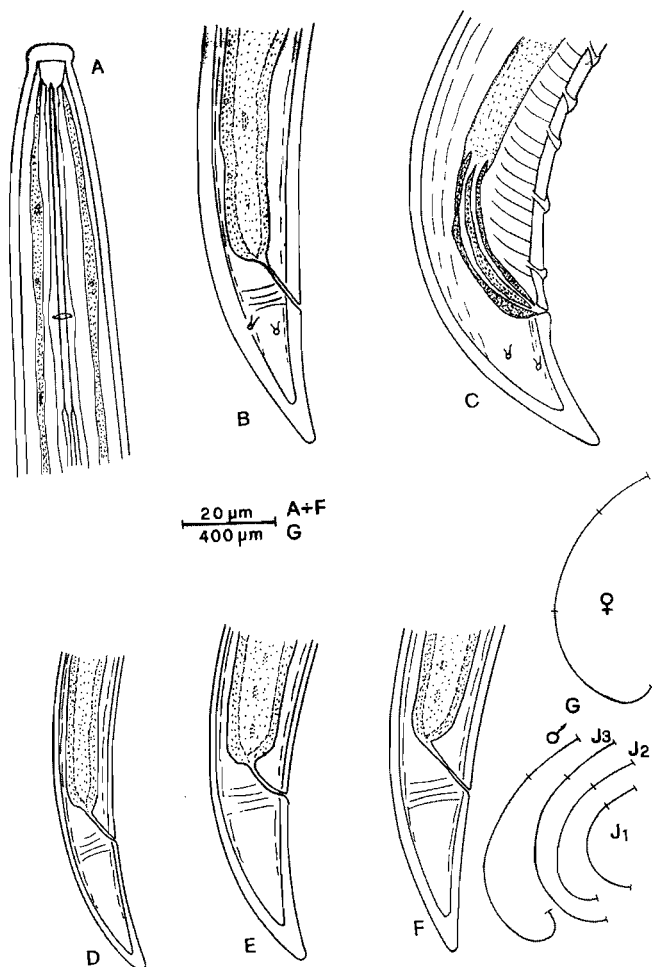
Juveniles occurred in the populations collected at Eustis and Quincy and three stages were identifiable in both populations (Fig. 3).

Two males, occurred in each of the populations from Yancey and Merrit Island. They are similar to females and the adanal pair of supplements is preceded by a row of nine or eleven ventromedian supplements.

*Xiphinema citricolum* differs from *X. americanum* Cobb, 1913 (Lamberti and Golden, 1984) in its more set off and frontally flattened lip region (only slightly

**Table I.** Populations of *Xiphinema* from Florida studied (+).

Population number Fl. n	Locality	Host	Studies performed			Species identification
			Biometrics	Isozymes	DNA	
124	Lake Alfred	Live oak, <i>Quercus virginiana</i> Mill.	+			<i>X. laevistriatum</i>
124 bis	Lake Alfred	Live oak	+	+		<i>X. georgianum</i>
156	Labelle	Long needle pine, <i>Pinus palustris</i> Mill.	+	+	+	<i>X. laevistriatum</i>
158	Labelle	<i>Citrus</i> sp.	+	+		<i>X. laevistriatum</i>
183	Moore Haven	Australian pine, <i>Casuarina</i> sp.	+	+	+	<i>X. laevistriatum</i>
192	Oklawaha	Swingle citrumelo <i>Citrus paradisi</i> Macf. x <i>Poncirus trifoliata</i> (L.) Raf.	+	+		<i>X. citricolum</i>
201	Altoona	Sour orange, <i>Citrus aurantium</i> L.	+	+	+	<i>X. floridae</i>
201 bis	Altoona	Sour orange	+			<i>X. citricolum</i>
205	Altoona	Sour orange	+	+	+	<i>X. citricolum</i>
218	Eustis	Cleopatra mandarin, <i>Citrus reticulata</i> Blanco	+	+	+	<i>X. citricolum</i>
227	Quincy	Bermuda grass, <i>Cynodon dactylon</i> (L.) Pers.	+	+	+	<i>X. citricolum</i>
275	Merrit Island	Sea grape, <i>Coccoloba uvifera</i> (L.) Jacq	+			<i>X. citricolum</i>
284	Merrit Island	<i>Casuarina</i> sp.	+	+	+	<i>X. laevistriatum</i>

**Fig. 1.** *Xiphinema citricolum*: A, female anterior region; B, female posterior region; C, male posterior region; D-F, posterior region of first, second and third juvenile stage respectively; G, habitus.

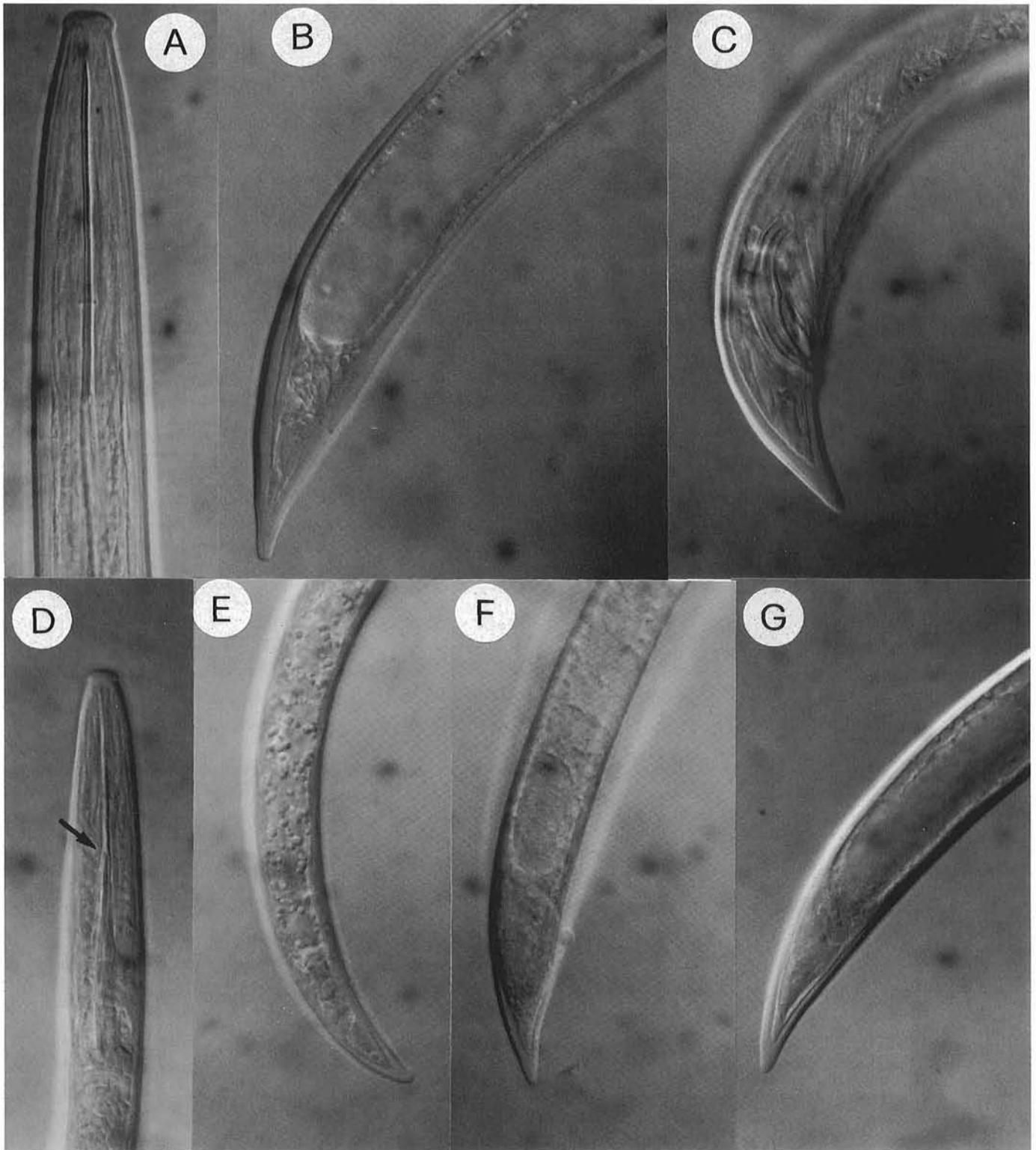
separated from the rest of the body and frontally rounded in *X. americanum*), straight, symmetrical, acute tail (dorsally curved and subacute terminus in *X. americanum*) and longer hyaline portion of the tail ( $J = 7 \mu\text{m}$  in *X. americanum*)

*Xiphinema citricolum* is similar to *X. pachtaicum* (Tulaganov, 1938) Kirjanova, 1951; *X. lambertii* Bajaj et Jairajpuri, 1977; *X. intermedium* Lamberti et Bleve Zacheo, 1979; *X. oxycaudatum* Lamberti et Bleve Zacheo, 1979; *X. peruvianum* Lamberti et Bleve Zacheo, 1979; *X. tarjanense* Lamberti et Bleve Zacheo, 1979; *X. tenuicutis* Lamberti et Bleve Zacheo, 1979; *X. bricolense* Ebsary, Vrain et Graham, 1989 and *X. pakistanense* Nasira et Maqbool, 1998.

However, compared to *X. pachtaicum* (Lamberti and Bleve-Zacheo, 1979), *Xiphinema citricolum* has lower  $c$  value (always more than 55 in *X. pachtaicum*), anterior vulva (V more than 55% in *X. pachtaicum*), more expanded lip region (diam. at lip region ca.  $8 \mu\text{m}$  in *X. pachtaicum*) and straight tail (ventrally slightly bent and concave in *X. pachtaicum* according to Lamberti and Martelli, 1971); compared to *X. lambertii* (Bajaj et Jarajpuri, 1977) *X. citricolum* has longer body ( $L = 1.3-1.4$  in *X. lambertii*), longer odontostyle ( $55-64 \mu\text{m}$  in *X. lambertii*), posterior basal guide ring ( $48-54 \mu\text{m}$  from the anterior extremity in *X. lambertii*) and straight tail (ventrally bent and concave in *X. lambertii*); compared to *X. intermedium* (Lamberti and Bleve-Zacheo, 1979), *X. citricolum* has higher  $a$  and  $c'$  values ( $a = 43$  and  $c' = 1.5$  in *X. intermedium*) and straight pointed tail (ventrally bent and concave in *X. intermedium*); compared to *X. oxycaudatum* (Lamberti and Bleve Zacheo, 1979), *X. citricolum* has more expanded lip region (slightly set off in *X. oxycaudatum*), longer hyaline portion of the tail ( $J = 9$

Table II. Morphometrics (mean  $\pm$  standard deviation; range) of *X. citricolum*.

Locality (populations)	Eustis (Fl. 218)				Quincy (Fl. 227)				Altoona (Fl. 205)		Merrit Island (Fl. 275)	
	10 ♀	3 J <sub>1</sub>	10 J <sub>2</sub>	15 J <sub>3</sub>	10 ♀	5 J <sub>1</sub>	20 J <sub>2</sub>	20 J <sub>3</sub>	10 ♀	1 ♂	10 ♀	1 ♂
n												
L (mm)	1.6 $\pm$ 0.12 1.5-1.8	0.72 $\pm$ 1.67 0.71-0.74	0.88 $\pm$ 4.04 0.82-0.94	1.1 $\pm$ 0.06 1.1-1.3	1.5 $\pm$ 0.06 1.4-1.6	0.68 $\pm$ 1.30 0.67-0.70	0.87 $\pm$ 4.61 0.76-0.94	1.2 $\pm$ 0.09 1.1-1.3	1.5 $\pm$ 0.05 1.5-1.6	1.5	1.4 $\pm$ 0.05 1.4-1.5	1.4
a	51 $\pm$ 2.95 46.4-54.7	37.4 $\pm$ 1.65 35.7-39	40 $\pm$ 1.59 37.2-41.6	43.7 $\pm$ 1.92 39.4-46.4	48.6 $\pm$ 1.99 45.7-51	36.8 $\pm$ 2.71 32.8-40	40 $\pm$ 1.58 37.2-42.8	44.2 $\pm$ 2.18 40.5-49	49 $\pm$ 3.05 45.3-54.4	47.2	47.7 $\pm$ 2.60 44-51.7	45.7
b	6 $\pm$ 0.46 5.4-7	4.2 $\pm$ 0.25 3.9-4.4	4.4 $\pm$ 0.37 4-5	4.7 $\pm$ 0.41 4.1-5.8	5.3 $\pm$ 0.29 5-5.7	4 $\pm$ 0.33 3.8-4.6	4.3 $\pm$ 0.37 3.7-5.6	4.9 $\pm$ 0.41 4.2-5.8	6 $\pm$ 0.47 5.5-7	5.7	5.1 $\pm$ 0.31 4.7-5.8	5.0
c	48.3 $\pm$ 4.01 42.5-56.6	22 $\pm$ 1.60 20.3-23.5	25.4 $\pm$ 1.44 23-28.2	33 $\pm$ 1.81 28-35.6	44.8 $\pm$ 1.38 43.2-46.7	23.5 $\pm$ 0.42 23-24	26.8 $\pm$ 1.18 25-29	33.8 $\pm$ 3.12 29.7-38.8	47 $\pm$ 3.62 40-52.3	44.8	43 $\pm$ 2.40 38.3-46.4	41
c'	1.8 $\pm$ 0.09 1.7-2	2.7 $\pm$ 0.06 2.6-2.7	2.5 $\pm$ 0.11 2.4-2.7	2.2 $\pm$ 0.11 2-2.3	1.8 $\pm$ 0.09 1.6-1.9	2.5 $\pm$ 0.07 2.4-2.6	2.4 $\pm$ 0.08 2.3-2.5	2.1 $\pm$ 0.09 1.9-2.2	1.7 $\pm$ 0.09 1.6-1.8	1.6	1.8 $\pm$ 0.10 1.7-2	1.5
V	52 $\pm$ 1.26 50-53	---	---	---	52 $\pm$ 1.07 50-53	---	---	---	51.7 $\pm$ 1.58 50-53	---	52 $\pm$ 0.92 51-53	---
Odontostyle $\mu$ m	78.6 $\pm$ 1.53 76.5-81.8	39.2 $\pm$ 1.73 38.2-41.2	48 $\pm$ 1.14 46-49.4	60.5 $\pm$ 1.39 57.6-63.5	74 $\pm$ 2.58 70-79.4	38.7 $\pm$ 1.98 36.5-41.2	45.8 $\pm$ 1.50 43.5-48.2	59.3 $\pm$ 2.36 55.3-63	79.2 $\pm$ 2.53 75.3-83	82.3	78.2 $\pm$ 2.24 74.7-80.6	81.2
Odontophore $\mu$ m	47 $\pm$ 1.58 44-48.8	27.6 $\pm$ 0.98 26.5-28.2	33.8 $\pm$ 1.60 31.2-36.5	41 $\pm$ 1.75 38.2-43.5	45.2 $\pm$ 1.54 41.8-47.6	28.4 $\pm$ 1.00 27-29.4	34.5 $\pm$ 1.27 32.3-36.5	41 $\pm$ 1.51 38-43.5	46.8 $\pm$ 1.54 44-48.8	48.8	45.4 $\pm$ 1.49 44-47.6	42.3
Replacement odontostyle $\mu$ m	---	47.2 $\pm$ 0.35 47-47.6	60.7 $\pm$ 2.71 56-66	79.9 $\pm$ 2.25 74.7-82.3	---	46.2 $\pm$ 0.63 45.3-47	60.4 $\pm$ 2.27 56-64	76.4 $\pm$ 3.08 68.8-82	---	---	---	---
Oral aperture to basal guide ring $\mu$ m	65.8 $\pm$ 1.56 62.3-68.8	31.4 $\pm$ 0.69 30.6-31.8	40 $\pm$ 1.07 38.2-42.3	51.5 $\pm$ 1.77 48.2-55.3	61.8 $\pm$ 2.11 57-64.7	29.5 $\pm$ 0.27 29.4-30	38.5 $\pm$ 1.00 37-41.2	50 $\pm$ 1.76 47.6-53	65.5 $\pm$ 2.30 62.3-68.8	68.8	63.4 $\pm$ 1.66 60.6-66	64.7
Tail $\mu$ m	33.2 $\pm$ 1.94 29.4-35.3	34 $\pm$ 0.69 33.5-34.7	35 $\pm$ 1.69 32.3-37	35 $\pm$ 1.70 32.3-38.8	33 $\pm$ 1.70 30-35.3	29.3 $\pm$ 0.89 28.2-30.6	32.7 $\pm$ 1.27 30.6-34.7	34.7 $\pm$ 1.50 31.8-37	32.2 $\pm$ 1.76 29.4-34.7	33.5	33.6 $\pm$ 1.69 31.8-36.5	34.1
J (hyaline portion of tail) $\mu$ m	11.2 $\pm$ 1.08 10-13.5	3.5 $\pm$ 0.55 3-4.1	5.9 $\pm$ 0.73 4.7-7	7.8 $\pm$ 0.85 6-8.8	9.6 $\pm$ 1.15 8.8-11.8	3.6 $\pm$ 0.27 3.5-4.1	5.1 $\pm$ 0.44 4.1-6	6.8 $\pm$ 0.73 6-8.2	10.6 $\pm$ 0.87 9.5-12	10.6	10.5 $\pm$ 0.60 10-11.2	10
Body diam. at lip region $\mu$ m	10.8 $\pm$ 0.31 10.6-11.2	7.6 $\pm$ 0.00 7.6-7.6	8.7 $\pm$ 0.25 8.2-8.8	9.5 $\pm$ 0.26 9.4-10	10.9 $\pm$ 0.42 10.6-11.8	7.7 $\pm$ 0.27 7.6-8.2	8.4 $\pm$ 0.27 8.2-8.8	9.2 $\pm$ 0.28 8.8-9.4	10.8 $\pm$ 0.42 10.6-11.8	11.8	10.9 $\pm$ 0.32 10.6-11.2	10.6
Body diam. at guide ring $\mu$ m	24.4 $\pm$ 0.80 23.5-26	13.7 $\pm$ 0.58 13-14	16.8 $\pm$ 0.45 16-17.6	20.3 $\pm$ 0.99 19.4-22.3	22.4 $\pm$ 0.64 21.8-23.5	13.3 $\pm$ 0.84 11.8-14	16.6 $\pm$ 0.41 16-17.6	19.8 $\pm$ 0.71 18.2-20.6	24.6 $\pm$ 0.89 23-26	24.7	23.6 $\pm$ 0.85 22.3-24.7	23.5
Body diam. at base of oesophagus $\mu$ m	29.4 $\pm$ 1.39 27.6-31.8	17.8 $\pm$ 1.39 17-19.4	20.5 $\pm$ 1.41 18.8-23	25 $\pm$ 1.83 23-29.4	27.7 $\pm$ 0.87 26-29.4	16.8 $\pm$ 1.27 15.3-18.8	20.4 $\pm$ 0.74 19.4-21.8	24.7 $\pm$ 1.42 22.3-26.6	29 $\pm$ 1.54 26.5-31.8	28.8	28 $\pm$ 1.28 26-30	26
Body diam. at mid-body or vulva $\mu$ m	31.4 $\pm$ 1.05 29.4-33	19.4 $\pm$ 1.04 18.8-20.6	22 $\pm$ 1.79 20.6-25.3	26.6 $\pm$ 1.88 24.7-30.6	30.5 $\pm$ 1.39 28.2-32.3	18.6 $\pm$ 1.24 17.6-20.6	21.8 $\pm$ 0.97 20.6-23.5	26.5 $\pm$ 1.44 24-28.8	31.6 $\pm$ 2.03 29.4-35.3	31.8	30.2 $\pm$ 1.56 27-32.3	30.6
Body diam. at anus $\mu$ m	18 $\pm$ 0.93 17-19.4	12.8 $\pm$ 0.40 12.3-13	13.8 $\pm$ 0.73 13-15.3	16.3 $\pm$ 1.05 15.3-18.8	18.5 $\pm$ 1.14 17.6-21.2	11.6 $\pm$ 0.33 11.2-11.8	13.7 $\pm$ 0.48 13-14.7	16.7 $\pm$ 0.81 14.7-17.6	18.8 $\pm$ 1.27 17-21.2	20.6	18.5 $\pm$ 1.01 16.5-19.4	22.3
Body diam. at beginning of J $\mu$ m	7.4 $\pm$ 0.42 7-8.2	4.1 $\pm$ 0.00 4.1-4.1	4.4 $\pm$ 0.51 4.1-5.3	5.6 $\pm$ 0.47 5.3-6.5	7.4 $\pm$ 0.58 7-8.8	3.9 $\pm$ 0.33 3.5-4.1	4.5 $\pm$ 0.38 4.1-5.3	5.8 $\pm$ 0.34 5.5-6.5	7.6 $\pm$ 0.52 7-8.2	7	7.2 $\pm$ 0.48 6.5-8.2	8.2
Spicules $\mu$ m	---	---	---	---	---	---	---	---	---	44	---	41.2



**Fig. 2.** Photomicrographs of *X. citricolum*: A, female anterior region; B, female posterior region; C, male posterior region; D, first stage juvenile anterior region (arrow indicates the tip of the replacement odontostyle inserted in the odontophore); E-G, posterior region of first, second and third juvenile stage respectively.

$\mu\text{m}$  in *X. oxycaudatum*) and straight more symmetrical and more gradually tapering tail (dorsally convex, asymmetrical in *X. oxycaudatum*); compared to *X. peruvianum* (Lamberti and Bleve Zacheo, 1979), *X. citricolum* has lower  $c$  value (56 in *X. peruvianum*), higher  $c'$  value (1.4 in *X. peruvianum*), longer hyaline portion of

the tail ( $J=8$  in *X. peruvianum*) and straight symmetrical tail (dorsally convex, asymmetrical in *X. peruvianum*); compared to *X. tarjanense* (Lamberti and Bleve Zacheo, 1979), *X. citricolum* has longer body ( $L=1.3$  mm in *X. tarjanense*) and anterior vulva ( $V=54\%$  in *X. tarjanense*); compared to *X. tenuicutis* (Lamberti and

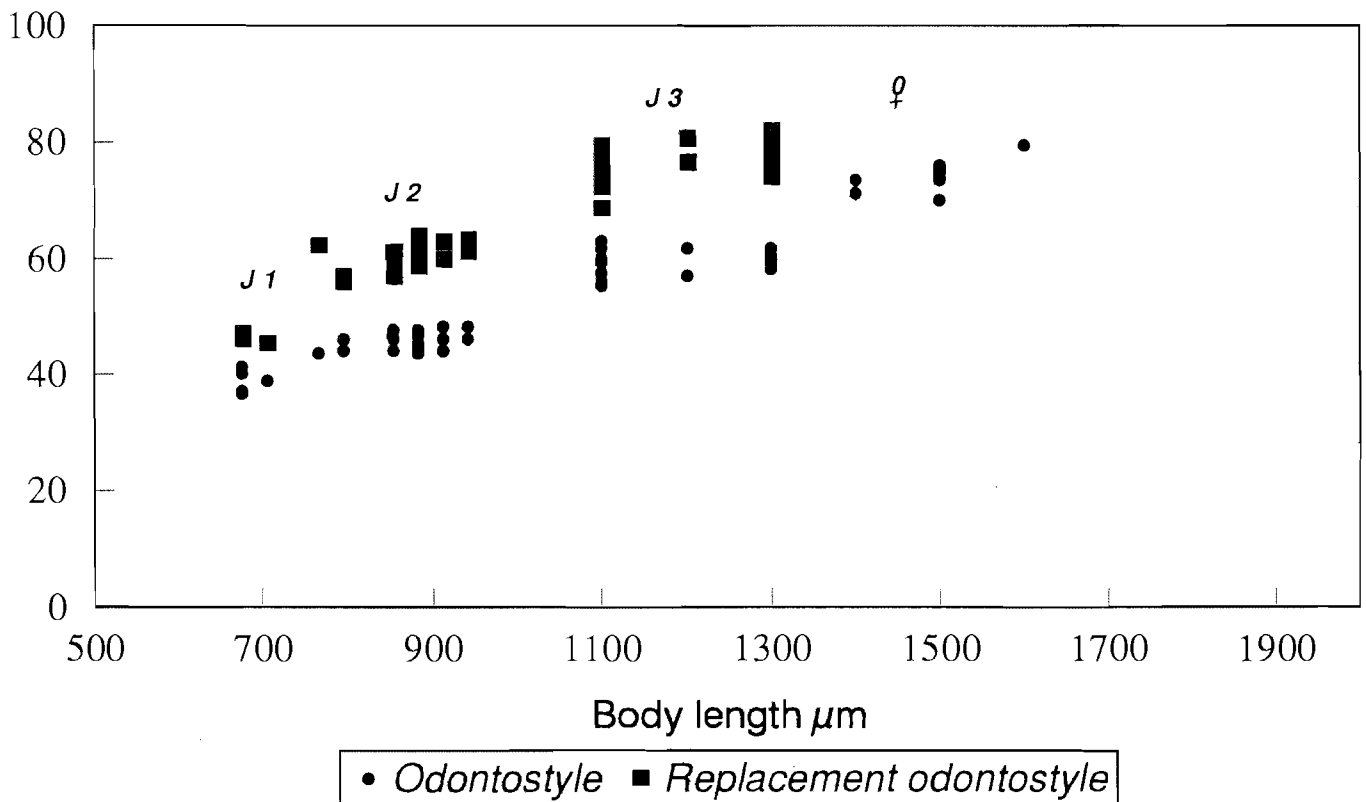
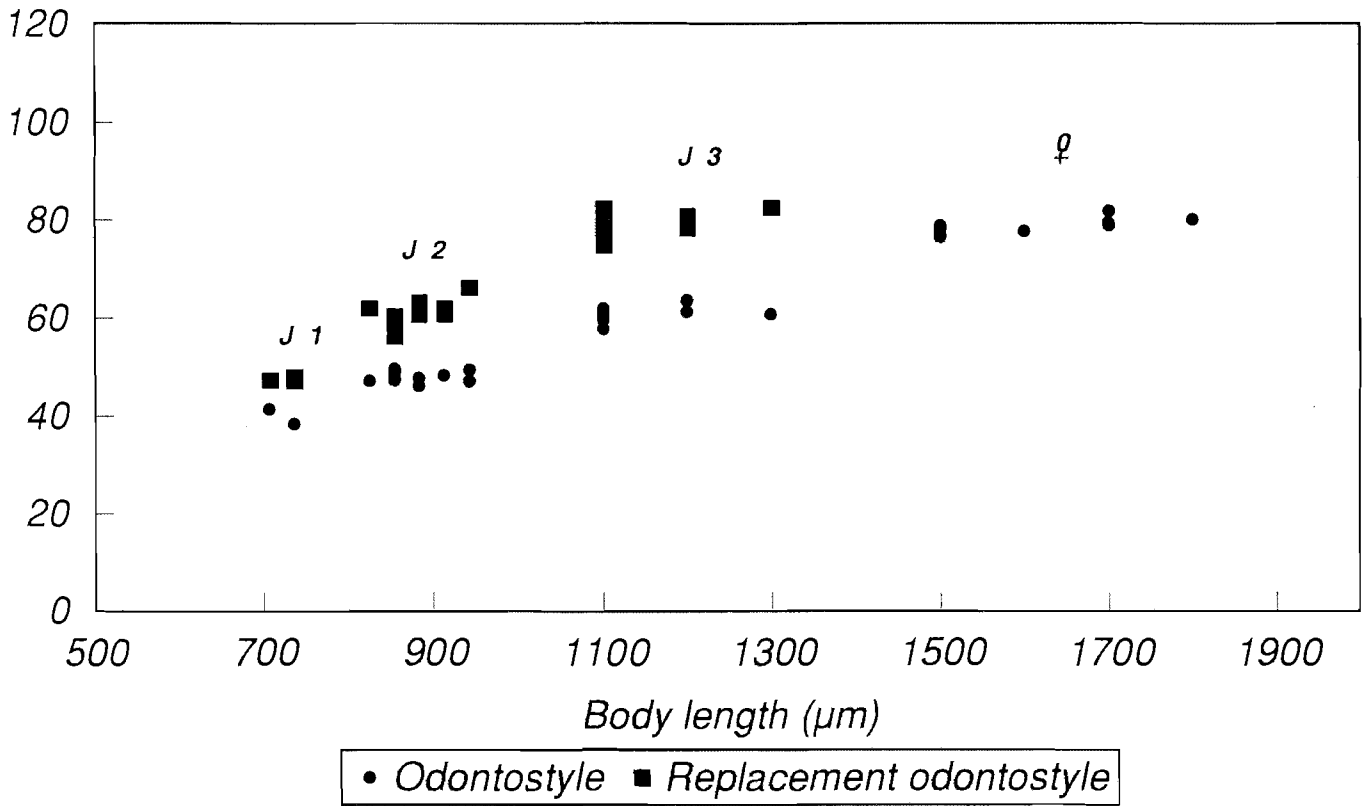


Fig. 3. Scatter diagrams plotting body and odontostyle length of individual juveniles and females of *X. citricolum* (top pop. Fl. 218, bottom pop. Fl. 227).

Bleve Zacheo, 1979), *X. citricolum* has lower c value (61 in *X. tenuicutis*) and longer tail (29 µm in *X. tenuicutis*) and hyaline portion of the tail (J= 8 in *X. tenuicutis*);

compared to *X. bricolense* (Ebsary *et al.*, 1989), *X. citricolum* has shorter body (L= 1.9 in *X. bricolense*); more expanded lip region (only moderately set off in *X.*

*bricolense*) and more gradually tapering tail; finally, compared to *X. pakistanense* (Nasir and Maqbool, 1998), *X. citricolum* has higher *c'* value (1.2 in *X. pakistanense*), longer odontostyle (66  $\mu\text{m}$  in *X. pakistanense*), posterior basal guide ring (56  $\mu\text{m}$  from the anterior extremity in *X. pakistanense*), more clearly set off lip region (moderately in *X. pakistanense*) and longer tail (29  $\mu\text{m}$  in *X. pakistanense*).

Leone *et al.*, (1997) consider *X. intermedium* the casual agent of a decline of Bermuda grass in Florida. This population should now be regarded as *X. citricolum*.

#### XIPHINEMA FLORIDAE

Lamberti *et Bleve-Zacheo*, 1979

(Tables I, III and VI; Figs 4, 5, 9 and 10)

A population of *X. floridae* occurred in the rhizosphere of sour orange near Altoona. Its morphometrics correspond to the original description with the only exception of being slightly longer (Lamberti and Bleve-Zacheo, 1979).

Table III. Morphometrics of *X. floridae*.

Locality	Altoona (pop. Fl. 201)
n	7 ♀
L (mm)	2±0.05 1.9-2
a	49.8±1.65 47.3-52.3
b	6.4±0.63 5.6-7.5
c	59.2±5.09 50.5-64.6
<i>c'</i>	1.4±0.05 1.3-1.4
V	52±1.21 50-54
Odontostyle $\mu\text{m}$	94.4±1.59 92.3-96.5
Odontophore $\mu\text{m}$	59.2±1.28 57.6-61.2
Oral aperture to basal guide ring $\mu\text{m}$	79±1.81 76.5-82.3
Tail $\mu\text{m}$	33±2.02 30-35.3
J (hyaline portion of tail) $\mu\text{m}$	7±0.32 6.5-7.6
Body diam. at lip region $\mu\text{m}$	13.2±0.27 13-13.5
Body diam. at guide ring $\mu\text{m}$	31±0.47 30.6-31.8
Body diam. at base of oesophagus $\mu\text{m}$	36±0.50 35.3-37
Body diam. at vulva $\mu\text{m}$	39.6±1.68 37.6-42.3
Body diam. at anus $\mu\text{m}$	24±0.64 23-24.7
Body diam. at beginning of J $\mu\text{m}$	9±0.32 8.8-9.4

Males and juveniles were not found.

*X. floridae* differs from *X. americanum* Cobb, 1913 (Lamberti and Golden, 1984) in its expanded lip region (separated by a shallow depression in *X. americanum*), longer body (L= 1.6 ca. in *X. americanum*), longer odontostyle (80  $\mu\text{m}$  ca. in *X. americanum*), conical straight tail (ventrally bent in *X. americanum*) and thicker body profile.

*X. floridae* is similar to *X. californicum* Lamberti *et Bleve-Zacheo*, 1979; *X. citricolum* Lamberti *et Bleve-Zacheo*, 1979; *X. oxycaudatum* Lamberti *et Bleve-Zacheo*, 1979; *X. peruvianum* Lamberti *et Bleve-Zacheo*, 1979; and *X. franci* Heyns *et Coomans*, 1994.

However, compared to *X. californicum* (Lamberti *et Bleve-Zacheo*, 1979), *X. floridae* has lower a value (60 in *X. californicum*), lower *c'* value (1.6 in *X. californicum*), wider tail and fatter body profile; compared to *X. citricolum* (Lamberti *et Bleve-Zacheo*, 1979), *X. floridae* has higher c value (48-50 in *X. citricolum*) and lower *c'* value (1.6-1.7 in *X. citricolum*); compared to *X. oxycaudatum* (Lamberti *et Bleve-Zacheo*, 1979), *X. floridae* has

Table IV. Morphometrics of *X. georgianum*.

Locality	Lake Alfred (pop. Fl. 124 bis)
n	10 ♀
L (mm)	2±0.11 1.8-2.1
a	47±1.64 44-49.6
b	5.9±0.46 5.4-6.8
c	73.3±4.26 66-79.2
<i>c'</i>	1.2±0.09 1.1-1.4
V	53±0.92 51-54
Odontostyle $\mu\text{m}$	118.7±3.00 113.5-125.3
Odontophore $\mu\text{m}$	59.5±2.54 54-61.8
Oral aperture to basal guide ring $\mu\text{m}$	96.5±3.38 88.8-100
Tail $\mu\text{m}$	27.3±1.66 24-30
J (hyaline portion of tail) $\mu\text{m}$	12.2±0.81 10.6-13.5
Body diam. at lip region $\mu\text{m}$	12.3±0.31 11.8-13
Body diam. at guide ring $\mu\text{m}$	32±1.44 30-34
Body diam. at base of oesophagus $\mu\text{m}$	38±2.01 35.3-41.2
Body diam. at vulva $\mu\text{m}$	42.6±2.58 38.8-47.6
Body diam. at anus $\mu\text{m}$	22.2±1.60 20.6-24.7
Body diam. at beginning of J $\mu\text{m}$	11.8±0.90 10.6-13.5

Table V. Morphometrics of *X. laevistriatum*.

Locality (populations)	Lake Alfred (pop. Fl. 124)	Labelle (pop. Fl. 156)	Labelle (pop. Fl. 158)	Moore Haven (pop. Fl. 183)	Merrit Island (pop. Fl. 284)			
					10 ♀	11 J <sub>1</sub>	16 J <sub>2</sub>	15 J <sub>3</sub>
<b>n</b>	10 ♀	10 ♀	10 ♀	10 ♀	10 ♀	11 J <sub>1</sub>	16 J <sub>2</sub>	15 J <sub>3</sub>
<b>L (mm)</b>	1.7±0.07 1.7-1.9	1.8±0.07 1.7-1.9	1.7±0.12 1.6-1.9	1.7±0.07 1.6-1.8	1.7±0.07 1.6-1.8	0.70±0.22 0.68-0.73	0.96±0.59 0.88-1.1	1.3±0.05 1.2-1.4
<b>a</b>	50.7±1.71 49-53.8	45.9±1.62 42.5-48	48±2.12 43.7-51.7	48±2.10 45.3-52.6	45±1.58 43-47	37.4±1.59 34.8-40.3	39.8±1.30 37.5-41.6	41.7±1.73 38.8-45
<b>b</b>	6±0.64 5.3-7.2	6.3±0.36 5.8-7	6.4±0.55 5.7-7.5	5.8±0.24 5.4-6	6.1±0.35 5.6-6.8	3.6±0.25 3.3-4.2	4.4±0.37 3.9-5	5.2±0.36 4.5-5.8
<b>c</b>	55.4±2.77 52.6-61.6	51.3±3.42 45.2-55.7	46±4.04 40-52	52.3±3.37 49-57.8	50.2±3.34 45.3-55.6	22.7±1.11 20.9-24.5	28±1.73 25-36	36.5±2.18 34-41.7
<b>c'</b>	1.7±0.08 1.5-1.8	1.6±0.09 1.5-1.8	1.7±0.13 1.6-2	1.5±0.11 1.3-1.6	1.5±0.03 1.4-1.5	2.6±0.10 2.5-2.8	2.2±0.09 2-2.3	1.8±0.11 1.6-2
<b>V</b>	52±1.32 49-53	51±1.16 50-53	51±1.20 50-53	53±1.85 48-55	51±1.15 50-53	---	---	---
<b>Odontostyle µm</b>	90±3.15 84.7-95.3	75.8±2.02 73.5-80.6	76.4±1.83 73-79.4	78.5±4.28 74.7-88.2	82±3.51 76.5-86.5	45±0.86 43.5-46	52±1.30 50-54	63.2±2.56 60-68.2
<b>Odontophore µm</b>	49.7±1.92 47-53	47.8±1.42 45.3-50	47.2±1.81 44-49.4	49.4±2.07 46-53	49.3±2.18 44.7-52.3	32±1.25 30-34	36.3±1.20 34.7-38.2	42.4±2.55 36.5-46.5
<b>Replacement odontostyle µm</b>	---	---	---	---	---	53±1.51 50.6-56	63±2.32 58.8-66.5	80.6±2.99 75.3-86.5
<b>Oral aperture to basal guide ring µm</b>	75±1.77 72.3-78.2	62±2.17 59-65.3	62.7±1.21 60.6-64.7	65.8±3.43 60-71.8	67.6±1.52 65.3-70.6	35±1.16 33-36.5	42.8±1.08 41.2-44	54±1.80 50-57
<b>Tail µm</b>	31±2.05 27.6-35.3	34.2±2.30 30.6-37.6	37.8±2.39 34-42.3	32±2.41 29.4-36.5	34±1.53 30.6-35.3	31±1.47 28.8-33.5	34.7±2.33 30.6-40	35.7±2.46 31.2-39.4
<b>J (hyaline portion of tail) µm</b>	8.1±0.68 7-8.8	12±0.99 10.6-13	11.3±1.23 9.4-13	7±0.66 6-8.2	9±0.67 8-10	5.2±0.74 4-6	5.4±0.72 4.7-7	6.3±0.49 5-7
<b>Body diam. at lip region µm</b>	8.9±0.25 8.8-9.4	11.4±0.29 11.2-11.8	11±0.42 10.6-11.8	11.2±0.40 10.6-11.8	11.4±0.31 11.2-11.8	8±0.31 7.6-8.2	9±0.31 8.8-9.4	10.4±0.37 10-11.2
<b>Body diam. at guide ring µm</b>	23.7±0.79 23-25.3	25.7±0.75 24.7-27	25.2±0.88 24-27	25.3±1.07 23.5-26.5	27±0.79 26-28.8	15.5±0.57 14.7-16.5	18.7±0.57 17.6-19.4	22.8±0.62 21.8-23.5
<b>Body diam. at base of oesophagus µm</b>	29±0.75 28.2-30.6	34±0.90 33-36	32.8±1.68 30.6-35.3	32.2±1.63 29.4-34.7	33.4±1.23 31.2-35.3	17.4±0.30 17-17.6	22.7±1.58 20-26	28.6±1.32 26.5-31.2
<b>Body diam. at mid-body or vulva µm</b>	34±1.00 31.8-35.3	38.2±1.66 36.5-41.8	36.3±2.78 33-41.2	35±1.65 32.3-37.6	37.8±1.74 35.3-41.3	19±0.32 18.2-19.4	24.3±1.62 21.2-27	31.2±1.74 27.6-33.5
<b>Body diam. at anus µm</b>	18.8±1.00 17.6-20.6	21.3±1.11 19.4-23	21.7±0.67 20.6-23	21.6±1.12 19.4-23.5	22.8±1.14 20-23.5	11.7±0.47 10.6-12.3	15.8±1.00 14-17	20.2±1.42 18.2-23.5
<b>Body diam. at beginning of J µm</b>	8.3±0.46 7.5-8.8	9.6±0.81 8.8-10.6	9±0.47 8.2-9.4	8.5±0.51 7.6-9.4	9.4±0.89 8.2-10.6	4.9±0.30 4.7-5.3	5.4±0.61 4.7-6.5	6.9±0.56 6-8.2

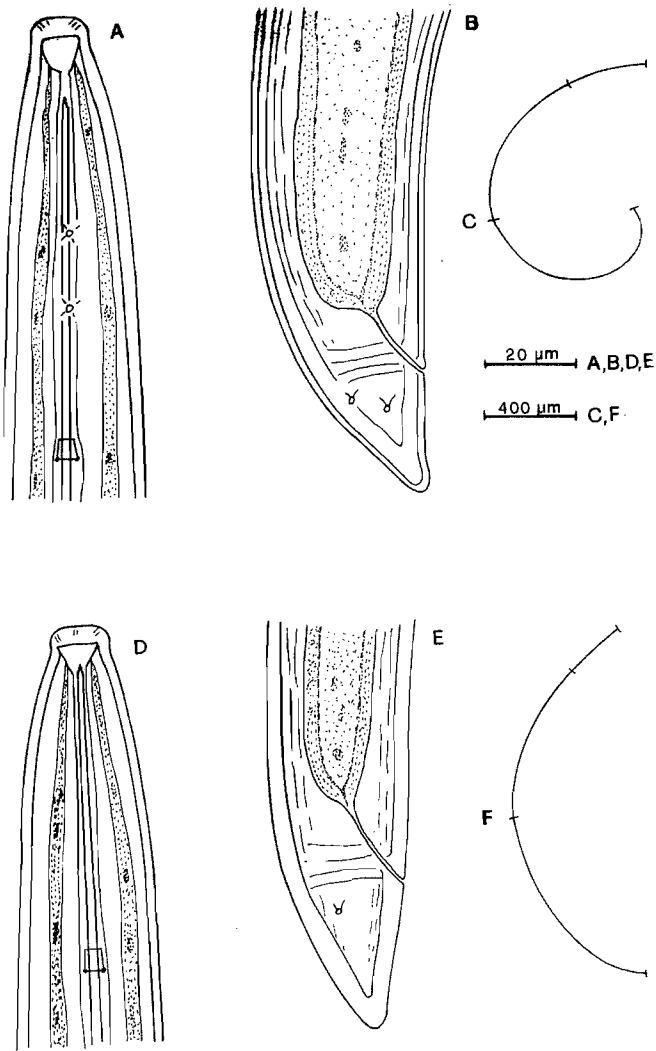


Fig. 4. Female of *Xiphinema georgianum*: A, anterior region; B, posterior region; C, habitus; female of *X. floridae*: D, anterior region; E, posterior region; F, habitus.

longer body ( $L=1.6$  in *X. oxycaudatum*), longer odontostyle ( $82-85\ \mu\text{m}$  in *X. oxycaudatum*) and wider tail; compared to *X. peruvianum* (Lamberti et Bleve-Zacheo, 1979), *X. floridae* has more expanded lip region (body diam at lip region  $10\ \mu\text{m}$  in *X. peruvianum*) and wider more symmetrical tail (subdigitate in *X. peruvianum*); finally, compared to *X. franci* (Heyns and Coomans, 1994), *X. floridae* has expanded lip region (almost continuous with the rest of the body in *X. franci*), longer body ( $L=1.4\ \text{mm}$  in *X. franci*) and wider straight tail (slightly bent and concave ventrally in *X. franci*).

#### *XIPHINEMA GEORGIANUM*

Lamberti et Bleve-Zacheo, 1979

(Tables I and IV; Figs 4, 5, 9 and 10)

A population of *X. georgianum* occurred in the rhizosphere of live oak at Lake Alfred. Its morphometrics fit the original description (Lamberti et Bleve-Zacheo, 1979) with the only exception of having slightly longer

odontostyle and odontophore.

Males and juveniles were not found.

*X. georgianum* differs from *X. americanum* Cobb, 1913 (Lamberti and Golden, 1984) in its longer body ( $L=1.6\ \text{mm}$  ca. in *X. americanum*), longer odontostyle ( $80\ \mu\text{m}$  ca. in *X. americanum*) lower  $c'$  value ( $1.8$  in *X. americanum*) and shorter tail ( $35\ \mu\text{m}$  ca. in *X. americanum*).

*X. georgianum* is similar to *X. paramonovi* Romanenko, 1981, *X. longistilum* Lamberti, Bravo, Agostinelli et Lemos, 1994 and *X. mesostilum* Lamberti, Bravo Agostinelli et Lemos, 1994.

However, compared to *X. paramonovi* (Romanenko, 1981), *X. georgianum* has longer odontostyle ( $103\ \mu\text{m}$  ca. in *X. paramonovi*), shorter tail ( $36\ \mu\text{m}$  ca. in *X. paramonovi*) and more tapering and pointed tail (rounded terminus in *X. paramonovi*); compared to *X. longistilum* (Lamberti et al., 1994), *X. georgianum* has shorter body ( $L=2.8\ \text{mm}$  in *X. longistilum*), anterior vulva ( $V=56$  in *X. longistilum*) and shorter tail ( $35\ \mu\text{m}$  in *X. longistilum*); finally, compared to *X. mesostilum* (Lamberti et al., 1994), *X. georgianum* possesses shorter body ( $L=2.5\ \text{mm}$  in *X. mesostilum*), anterior vulva ( $V=57$  in *X. mesostilum*) and much lower  $a$  and  $c$  values ( $91$  and  $98$  respectively in *X. mesostilum*).

#### *XIPHINEMA LAEVISTRITUM*

Lamberti et Bleve-Zacheo, 1979

(Tables I and VI; Figs 6-10)

Five populations were identified as *X. laevistriatum*. Their morphometrics are in the range of the original description (Lamberti et Bleve Zacheo, 1979); the only remarkable differences were observed in the population collected at Lake Alfred which, compared to the others, has longer odontostyle and posterior basal guide ring.

Juveniles occurred in the population from Merritt Island; males were not found.

*X. laevistriatum* differs from *X. americanum* Cobb, 1913 (Lamberti and Golden, 1994) in its slightly posterior vulva ( $V=50\%$  in *X. americanum*), lower value of  $c'$  ( $1.9$  in *X. americanum*) and longer hyaline portion of the tail ( $J=7\ \mu\text{m}$  in *X. americanum*).

*X. laevistriatum* is similar to *X. inaequale* (Khan et Ahmad, 1975) Khan et Ahmad, 1977; *X. luci* Lamberti et Bleve-Zacheo, 1979 and *X. minor* Ahmad, Lamberti, Rowat, Agostinelli et Srivastava, 1998.

However, compared to *X. inaequale* (Khan and Ahmad, 1975; Khan and Ahmad, 1977), *X. laevistriatum* has shorter body ( $L=2\ \text{mm}$  in *X. inaequale*), higher  $c'$  value ( $1.2$  in *X. inaequale*) and pointed terminus of the tail (rounded in *X. inaequale*); compared to *X. luci* (Lamberti and Bleve-Zacheo, 1979), *X. laevistriatum* has lower  $c$  value ( $65$  in *X. luci*), higher  $c'$  value ( $1.2$  in *X. luci*), shorter odontostyle ( $95\ \mu\text{m}$  in *X. luci*) and pointed tail terminus (rounded in *X. luci*); finally, compared to *X. minor* (Ahmad et al., 1998), *X. laevistriatum* has longer body ( $L=1.5\ \text{mm}$  in *X. minor*), longer odontostyle ( $69\ \mu\text{m}$  in *X. minor*) and longer tail ( $30\ \mu\text{m}$  in *X. minor*).



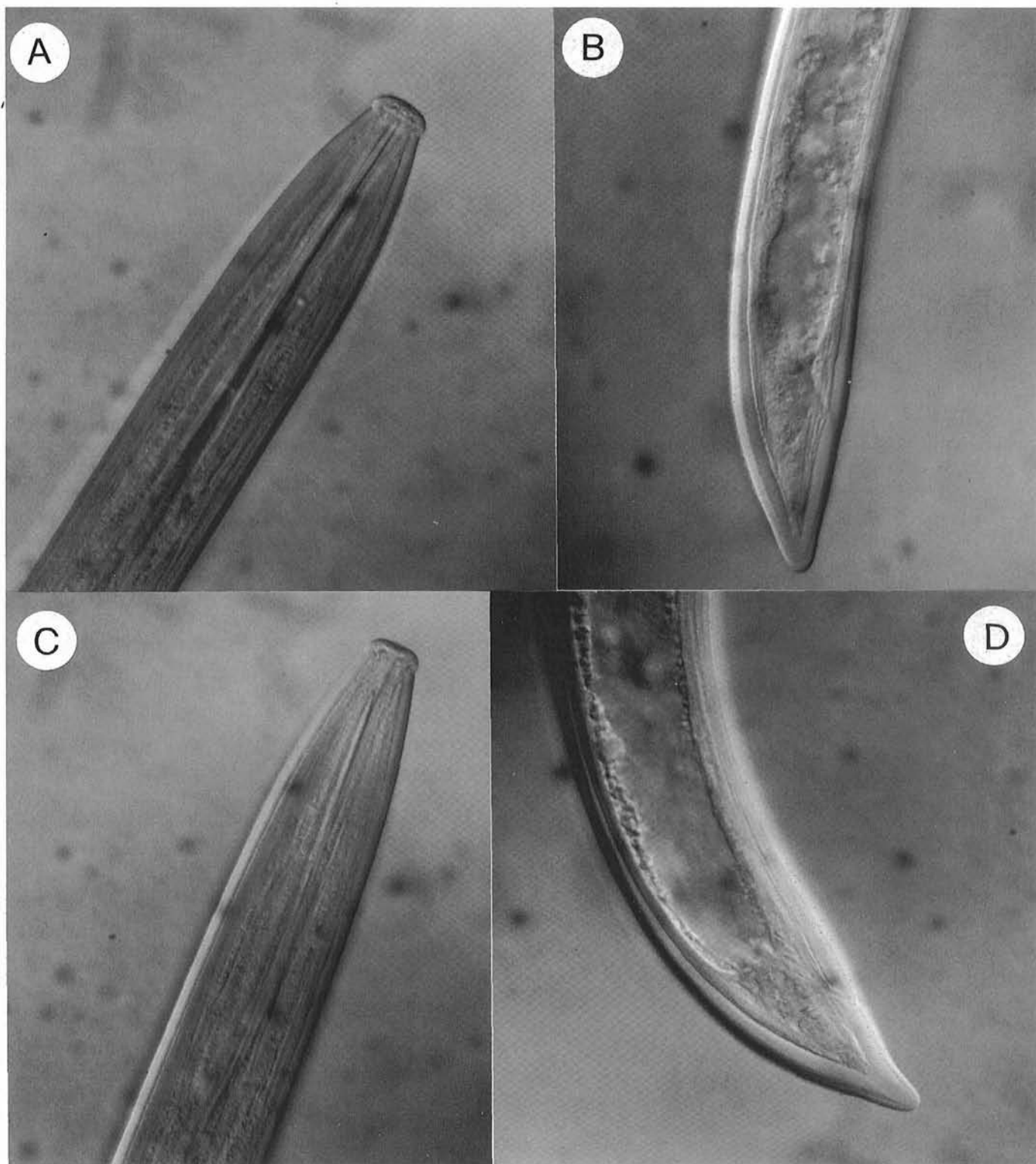


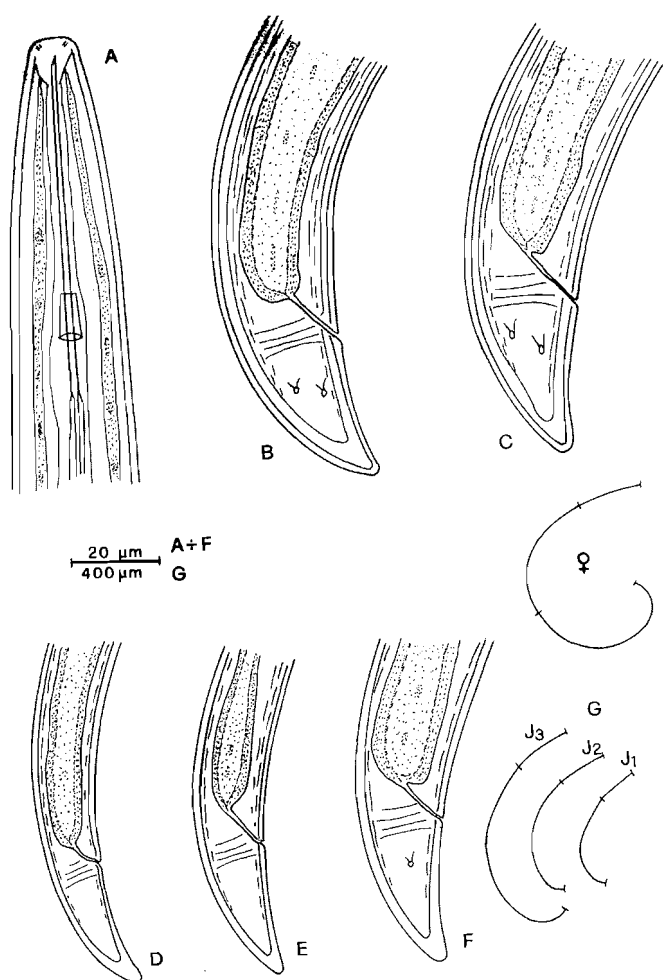
Fig. 5. Photomicrographs of females of *X. floridae*: A, anterior and B, posterior regions and *X. georgianum*: C, anterior and D, posterior regions.

Iso-electrofocusing of SOD isoforms (Fig. 9) grouped the populations into four phenotypes (Fig. 10): (1) population Fl. n. 192 from Oklawaha, identified as *X. citricolum*, which is characterized by a band of pH 7.1; (2) population Fl. n. 201 from Altoona, identified as *X. floridae* which shows only one band of SOD activity at basic pH; (3) a group comprehending populations Fl. n. 124 bis, from Lake Alfred, identified as *X. geor-*

*gianum*; Fl. n. 158 from Labelle, identified as *X. laevistriatum* and Fl. n. 205 from Altoona, identified as *X. citricolum*, characterized by three main bands at pH 8.5, 6.1 and 4.8; (4) a group of five populations, including Fl. n. 156 from Labelle (*X. laevistriatum*) Fl. n. 183 from Moore Haven (*X. laevistriatum*), Fl. n. 218 from Eustis (*X. citricolum*), Fl. n. 227 from Quincy (*X. citricolum*) and Fl. n. 284 from Merrit Island (*X. laevistria-*

**Table VI.** Estimated restriction fragment sizes (bp) of PCR amplified ITS of populations within the *Xiphinema americanum* group.

Enzymes	A				B		C	
	Pop. Fl. n. 156	Pop. Fl. n. 205	Pop. Fl. n. 218	Pop. Fl. n. 227	Pop. Fl. n. 201	Pop. Fl. n. 183	Pop. Fl. n. 284	
ND	1500	1500	1500	1500	1500	1500	1500	
<i>Alu</i> I	1100	(1200), (900), 750, 390, 140	730, 380	(1100), (850), 750, 380, 150	750, 380, 250, 130, 120	750, 390, 150	(1100), (850), 750, 380, 150	
<i>Bam</i> HI	780, 650	780, 650	800, 650	780, 650	uncut	780, 650	780, 650	
<i>Dde</i> I	430, 390, 380	420, 390, 370, 160	430, 380, 350, 150	430, 390, 380, 180	780, 360, 290, 150, 70	430, 390, 380, 180, 80	430, 390, 370, 180	
<i>Hinf</i> P I	560, 380, 300, 280, 200, 150	550, 390, 300, 250, 190, 150	550, 380, 320, 250, 200	(560), 380, 320, (290), (280), 200, 150	480, 320, 280, 200, 180, 120 110, 80	560, 380, 320, 290, 280, 200 150	560, 380, 320, 280, 200, 150	
<i>Rsa</i> I	900, 550	900, 550	900, 550	900, 550	900, 550	900, 550, 280, 250	900, 550, 280, 250	
<i>Xba</i> I	1350, 150	1350, 150	1350, 150	1350, 150	1350, 150	1350, 150	1350, 150	

**Fig. 6.** *Xiphinema laevistriatum*: A, female anterior region; B and C, female posterior region; D-F, posterior region of first, second and third juvenile stage respectively; G, habitus.

*tum*), varied from the group (3) for having an additional band at pH 5.6.

The DNA analysis, performed on seven populations

(Table VI; Fig. 11) indicated that the PCR product of the ITS region of three species within the *X. americanum*-group is 1.5 kb. The population of *X. georgianum* was not analyzed; populations Fl. n. 205, 218 and 227, morphometrically identified as *X. citricolum* (Table VI, A; Fig. 11A) showed the same restriction pattern, as population Fl. n. 156 which morphometrically was identified as *X. laevistriatum*; populations Fl. n. 183 and 284, identified as *X. laevistriatum*, were similar to *X. citricolum*, however, showed a peculiar pattern with the enzyme *Rsa* I (Table VI, C; Fig. 11C) which yielded two fragments of 280 and 250 bp, not present in *X. citricolum*; population Fl. n. 201 identified as *X. floridae*, was not digested by the enzyme *Bam* HI and showed a peculiar restriction profile with enzymes *Dde* I and *Hinf* I (Table VI, B; Fig. 11B).

## CONCLUSIONS

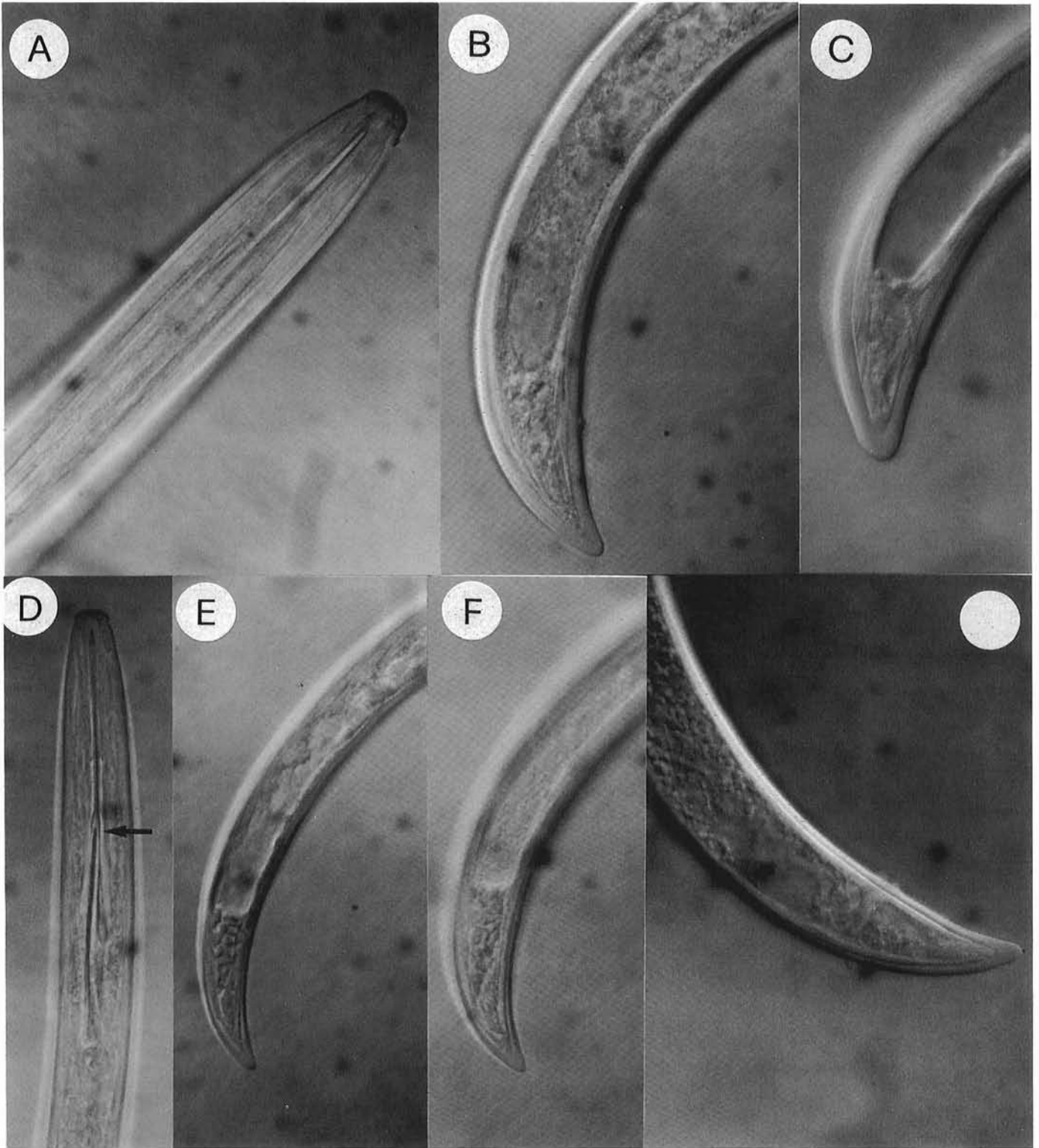
This work does not solve the complex and debated problem of the valid species belonging to the *X. americanum*-group. However, it permits to characterize four species which clearly possess discriminant differences and for which the following diagnoses are proposed.

*X. citricolum*, body length of ca. 1.6 mm, lip region set off from body profile, odontostyle length of ca. 80 µm; V= 52-54%, value of c' 1.6-1.8, tail straight, symmetrically conoid, with pointed terminus;

*X. floridae*, body length 1.8-2.0 mm, lip region set off from body profile, odontostyle length of ca. 90-95 µm; V= 50-52%, value of c' 1.1-1.4, tail straight, symmetrically conoid, with pointed terminus;

*X. georgianum*, body length of ca. 2 mm, lip region set off from body profile, odontostyle length of 110-120 µm, V ca. 53%, value of c' 1.2-1.3, tail straight, symmetrically conoid, with pointed terminus;

*X. laevistriatum* body length 1.7-1.8 mm, lip region continuous with body profile, odontostyle length of 75-



**Fig. 7.** Photomicrographs of *X. laevistriatum*: A, female anterior region; B and C, female posterior region; D, first stage juvenile anterior region (arrow indicates the tip of the replacement odontostyle inserted in the odontophore); E-G, posterior region of first, second and third juvenile stage respectively.

90  $\mu\text{m}$ ,  $V = 51-53\%$ , value of  $c' = 1.5-1.7$ , tail ventrally bent and slightly concave, with pointed terminus.

On the basis of our present knowledge SOD isoforms, isoelectrofocusing patterns did not discriminate between *X. americanum* populations identified by morphometric characters. It may be that small morphologi-

cal differences among *X. americanum*-group species are not likely to be resolved in a system based on enzyme polymorphism.

The DNA analysis revealed that more restriction enzymes are needed to characterize populations within the *X. americanum*-group.

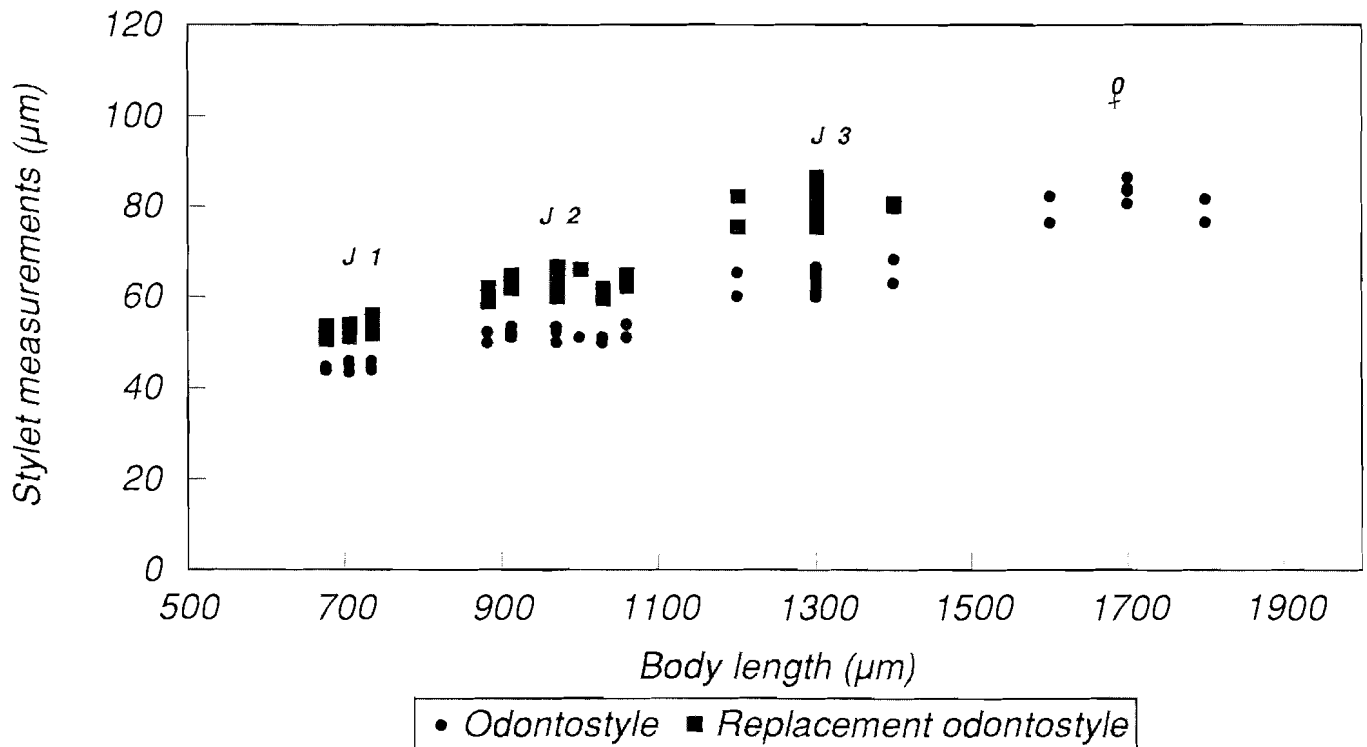
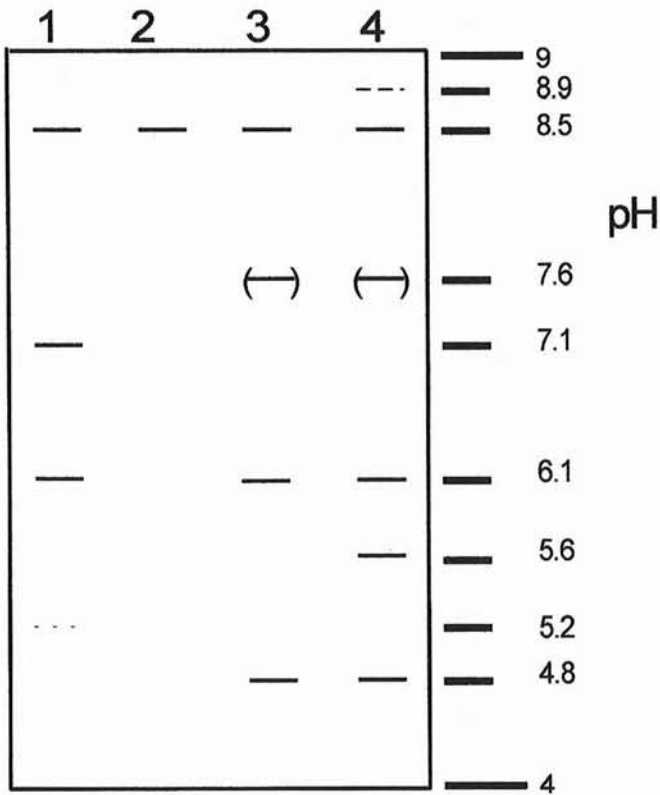


Fig. 8. Scatter diagram plotting body and odontostyle length of individual juveniles and females of *X. laevistriatum*.



Fig. 9. Isoelectrofocusing of SOD isozymes from extracts of populations belonging to the *Xiphinema americanum*-group. Mini-gels were stained for SOD, dried, scanned into computer images, turned into negatives and printed on photo quality paper. Bands of SOD activity appear black over a white background. Nematode populations are shown as follows: a) Fl 227; b) Fl 284; c) Fl 201; d) Fl 124 bis; e) Fl 183; f) Fl 218; g) Fl 205; h) Fl 192; i) Fl 158; l) Fl 156.



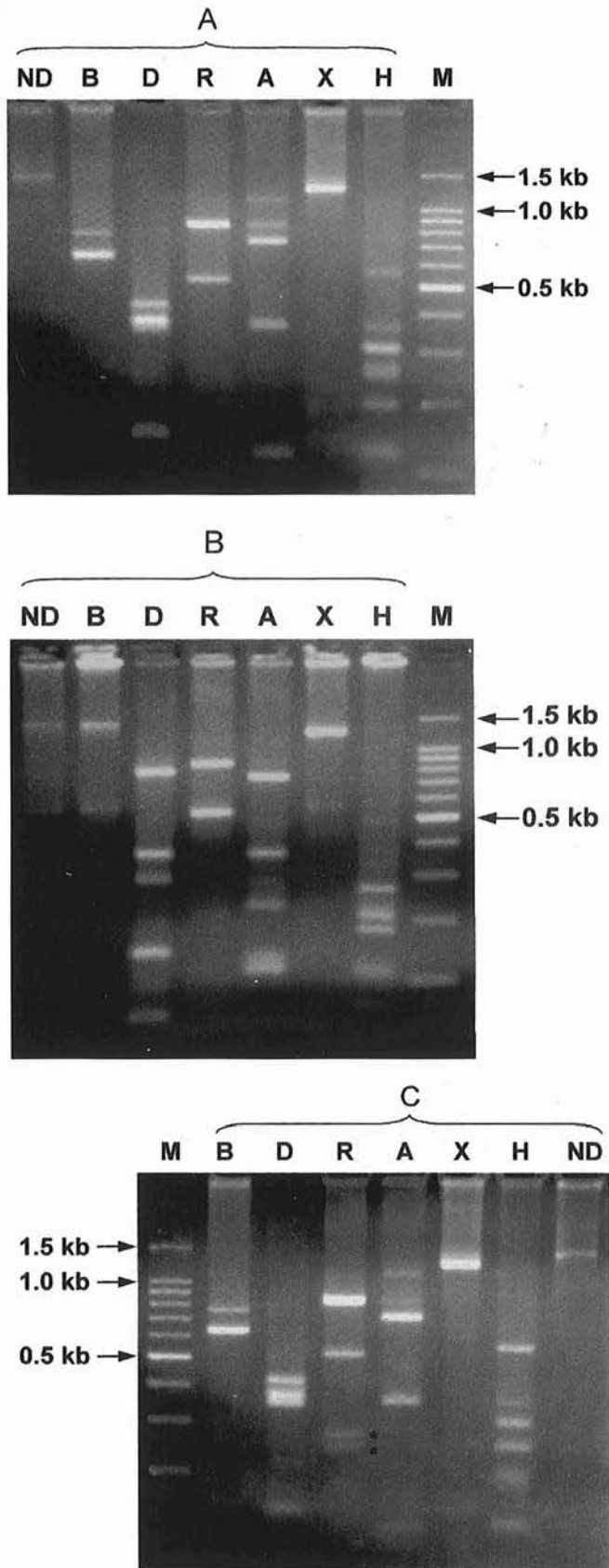
**Fig 10.** Diagram of the main 4 types of SOD isozyme patterns shown by the *X. americanum*-group populations. Type (1) includes Fl 192; type (2) Fl 201; type (3) Fl 124 bis, Fl 158, Fl 205; type (4) Fl 156, Fl 183, Fl 218, Fl 227, Fl 284.

#### ACKNOWLEDGEMENTS

The authors wish to express heartfelt gratitude to Teresa Lamberti who crossed the Atlantic with 30 Kg of soil samples in her hand-luggage, Jason Zellers for assistance in collecting and processing those samples, and Franco Zacheo for slide preparation.

#### LITERATURE CITED

- Ahmad M., Lamberti F., Rawat V.S., Agostinelli A. and Srivastava N., 1998 - Two new species within the *Xiphinema americanum*-group (Nematoda, Dorylaimida) from Garhwal Himalayas, India. *Nematologia Mediterranea*, 26: 131-138.
- Bajaj H.K. and Jairajpuri M.S., 1977 - Two new species of *Xiphinema* from India. *Nematologia Mediterranea*, 4 (1976): 195-200.
- Ebsary B.A., Vrain T.C. and Graham M.B., 1989 - Two new species of *Xiphinema* (Nematoda, Longidoridae) from British Columbia vineyards. *Canadian Journal of Zoology*, 67: 801-804.
- Heyns J. and Coomans A., 1994 - Four species of the *Xiphinema americanum* group (Nematoda, Longidoridae) from islands in the Western Indian Ocean. *Nematologica*, 40: 12-24.



**Fig. 11.** Restriction digests of the PCR amplification product of the ITS region of *X. americanum* - group populations: A, *X. citricolum* type; B, *X. floridiae* type; C, *X. laevistriatum* type, separated on a 2% agarose gel and stained with ethidium bromide (M= 100bp DNA ladder, ND= not digested, B= *Bam* HI, D= *Dde* I, R= *Rsa* I, A= *Alu* I, H= *Hinf* I, X= *Xba* I).

- Khan S.H. and Ahmad D.S., 1975 - Longidoroidea (Thorne, 1935) n. rank (Nematoda, Dorylaimina) with description of *Xiphinema neoamericanum* n. sp. from India and proposal for a new name for *Xiphinema americanum* sensu Carvalho (1956) non Cobb, 1913. *Nematologia Mediterranea*, 3: 23-28.
- Khan S.H. and Ahmad D.S., 1977. *Xiphinema inaequale* nom. nov. (syn. *Xiphinema neoamericanum* Khan et Ahmad, 1975). *Nematologia Mediterranea*, 5: 93.
- Lamberti F. and Bleve-Zacheo T., 1979 - Studies on *Xiphinema americanum* sensu lato with description of fifteen new species (Nematoda, Longidoridae). *Nematologia Mediterranea*, 7: 51-106.
- Lamberti F., Bravo M.A., Agostinelli A. and Lemos R.M., 1994 - The *Xiphinema americanum*-group in Portugal with description of four new species (Nematoda, Dorylaimida). *Nematologia Mediterranea*, 22: 189-218.
- Lamberti F. and Golden A.M., 1984 - Redescription of *Xiphinema americanum* Cobb, 1913 with comments on its morphometric variations. *Journal of Nematology*, 16: 204-206.
- Lamberti F. and Martelli G.P., 1971 - Notes on *Xiphinema mediterraneum* (Nematoda, Longidoridae). *Nematologica*, 17: 75-81.
- Lamberti F., Molinari S., Moens M. and Brown D.J.F., 2000 - The *Xiphinema americanum* group. I. Putative species, their geographical occurrence and distribution, and regional polytomous identification keys for the group. *Russian Journal of Nematology*, 8: 65-84.
- Lamberti F., Sabova M., De Luca F., Molinari S., Agostinelli A., Coiro M.I. and Valocka B., 1999 - Phenotypic variations and genetic characterization of *Xiphinema* populations from Slovakia (Nematoda, Dorylaimida). *Nematologia Mediterranea*, 27: 261-275.
- Leone A., Miano V., Lamberti F., Duncan L.W., Rich J.R. and Bleve-Zacheo T., 1997 - Cellular changes induced by *Xiphinema vulgare* in the roots of citrumele and by *Xiphinema intermedium* in the roots of Bermuda grass. *Nematologia Mediterranea*, 25: 199-207.
- Luc M., Coomans A., Loof P.A.A. and Baujard P., 1998 - The *Xiphinema americanum* group (Nematoda, Longidoridae). 2. Observations on *Xiphinema brevicollum* Lordello & Da Costa, 1961 and comments on the group. *Fundamental and Applied Nematology*, 21: 475-490.
- Molinari S., De Luca F., Lamberti F. and De Giorgi C., 1997 - Molecular methods for the identification of longidorid nematodes. *Nematologia Mediterranea*, 25: 55-61.
- Nasira K. and Maqbool M.A., 1998 - Description of *Xiphinema pakistanense* n. sp. and the male of *X. oxycaudatum* Lamberti & Bleve-Zacheo, 1979 with observations on *X. thornei* Lamberti & Golden, 1986 (Nematoda, Longidoridae) from Pakistan. *Pakistan Journal of Nematology*, 16: 1-12.
- Robbins R.T., 1993 - Distribution of *Xiphinema americanum* and related species in North America. *Journal of Nematology*, 25: 344-348.
- Romanenko N.D., 1981 - [A finding of a new species of nematode *Xiphinema paramonovi* n. sp. (Nematoda, Longidoridae) from the territory of the Soviet Union]. Tezisy Dokladov Pervoi Konferentsii po Nematodam Rastenii Pochvy i vod, pp. 68-69.
- Tarjan A.C., 1974. The dagger nematodes (*Xiphinema*, Cobb) of Florida. *Proceedings of the Soil and Crop Science Society of Florida*, 33: 92-95.