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NEMATICIDAL POTENTIAL OF SEED EXTRACTS: *IN VITRO* EFFECTS ON JUVENILE MORTALITY AND EGG HATCH OF *MELOIDOGYNE INCOGNITA* AND *M. JAVANICA*

by

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Summary. Seed extracts of *Acacia eburnea*, *Azadirachta indica*, *Cassia* sp., *Parkinsonia aculeata*, *Sesbania sesban* and *Poinciana regia* had the highest nematicidal potential against *Meloidogyne incognita* juveniles while maximum potential against *M. javanica* was shown by *A. eburnea*, *Cassia* sp., *Melia azedarach*, *S. sesban* and *Tribulus terrestris*, as indicated by strong activity in all concentrations. Similarly, *Calotropis procera* and *S. sesban* were the best in preventing egg hatch of *M. incognita*, and *C. procera*, *S. sesban* and *Chenopodium album* were the most effective against hatch of *M. javanica* eggs. *S. sesban* was outstanding in killing juveniles and preventing hatching of both the species.

Nematicidal properties of seed extracts have received less attention than those of other plant parts and seed cakes of some common oil-yielding plants (Prot and Kornprobst, 1983; Devakumar *et al.*, 1985; Lee, 1990). This study was undertaken after the encouraging results obtained for some seed extracts, against root knot nematodes, in an earlier study in this laboratory.

Materials and methods

The screening of thirty locally available seeds was carried out, *in vitro*, in 1993 for their nematicidal activity. Seeds of thirty (Table I) and twenty-seven (Table II) species, respectively, were tested against *Meloidogyne incognita* (Kofoid *et* White) Chitw. and *M. javanica* (Treub) Chitw. juveniles (J₂) and twenty-eight (Tables III and IV) for inhibitory effects on egg hatch of both species.

Seeds were ground in a grinder and immersed in 100 ml of distilled water for 24 hours, centrifuged at 4,000 rpm for 10 minutes and then filtered. Treatments were 1:10, 1:20 and 1:40 concentrations. Distilled water was used as the control.

Nematode populations were maintained on brinjal and okra in pot cultures. For mortality studies, 80-100 J₂/ml suspension were exposed to 10 ml of each test solution in 3 cm diameter Petri-dishes with three replicates of each treatment. The dishes were kept at 27 ± 1 °C in a B.O.D. incubator for 24 hours. Dead and live juveniles were then counted. Mortality was ascertained by transferring some of the treated, apparently dead juveniles to distilled water and examining them after 5-6 hours for any revival. Mortality was expressed as mean per cent.

For evaluating the effects on hatching, mature egg masses were separated from fresh roots. Four egg masses were placed in 10 ml of

TABLE I - In vitro screening of seed extracts for nematocidal activity against *Meloidogyne incognita* second stage juveniles (J₂).

Plant species	Family	Per cent J ₂ mortality, at 27 ± 1 °C, after 24 hrs			
		1:10	Dilutions		Control
			1:20	1:40	
<i>Acacia eburnea</i> Willd.	Leguminosae	100	98	91	0
<i>Azadirachta indica</i> A. Juss.	Meliaceae	100	95	89	0
<i>Cassia</i> sp.	Leguminosae	100	93	86	0
<i>Parkinsonia aculeata</i> L.	Leguminosae	98	92	84	0
<i>Sesbania sesban</i> (L.) Merr.	Leguminosae	98	91	81	0
<i>Eclipta alba</i> (L.) Hassk.	Compositae	98	84	41	0
<i>Acacia</i> sp.	Leguminosae	98	78	61	1
<i>Poinciana regia</i> Boj	Leguminosae	96	94	80	0
<i>Pongamia glabra</i> Vent. Jard. Malm.	Leguminosae	96	85	65	0
<i>Cassia fistula</i> L.	Leguminosae	96	72	44	1
<i>Melia azedarach</i> L.	Meliaceae	96	70	55	0
<i>Calotropis procera</i> R. Br.	Asclepiadaceae	95	80	62	0
<i>Terminalia bellerica</i> Roxb.	Combretaceae	92	82	49	0
<i>Withania somnifera</i> (L.) Dunal	Solanaceae	88	68	13	0
<i>Terminalia arjuna</i> (Roxb. ex DC) Wt. et Arn. Prodr.	Combretaceae	87	69	57	0
<i>Cassia occidentalis</i> L.	Leguminosae	86	66	11	2
<i>Parthenium hysterophorus</i> L.	Compositae	86	62	13	0
<i>Amaranthus viridis</i> L.	Amaranthaceae	85	42	11	0
<i>Chenopodium album</i> L.	Chenopodiaceae	83	55	34	0
<i>Nicotiana plumbaginifolia</i> Viv. Elench.	Solanaceae	68	31	9	0
<i>Croton bonplandianum</i> Baill.	Euphorbiaceae	62	43	17	0
<i>Tribulus terrestris</i> L.	Zygophyllaceae	54	32	12	0
<i>Euphorbia hirta</i> L.	Euphorbiaceae	43	18	6	0
<i>Malvastrum coromandelianum</i> ¹ (L.) Garcke	Malvaceae	29	15	10	0
<i>Cannabis sativa</i> L.	Cannabaceae	14	9	4	0
<i>Mimusops elengi</i> L.	Sapotaceae	11	2	0	0
<i>Lantana indica</i> L.	Verbenaceae	4	4	0	0
<i>Callistemon lanceolatus</i> DC. Prodr.	Myrtaceae	0	0	0	0
<i>Lagerstroemia speciosa</i> (L.) Pers.	Lythraceae	0	0	0	0
<i>Lagerstroemia thorelii</i> Gagnep.	Lythraceae	0	0	0	0

¹ With nematostatic effect.

each dilution in 3 cm diameter Petri-dishes. Distilled water served as the control. Each treatment was replicated three times. The dishes were kept at 27 ± 1 °C and emerged juveniles

were counted after 48 hours exposure. Mortality of emerged juveniles was also noted. Results were expressed as mean per cent hatch and mean per cent mortality.

Results

Nineteen seed extracts were highly toxic (83-100%) to *M. incognita* in the highest concentration (1:10) (Table I) and most of these also showed very good activity at 1:20 dilution. Mortality generally was much less at 1:40 concentration. However, *Acacia eburnea*, *Azadirachta indica*, *Cassia* sp., *Parkinsonia aculeata*, *Sesbania sesban* and *Poinciana regia* were highly effective (80-100%) in all dilutions.

Against *M. javanica*, seventeen seeds were highly active (at 1:10) and five were ineffective at all dilutions (Table II). *A. eburnea*, *Cassia* sp., *Melia azedarach*, *S. sesban* and *Tribulus terrestris* killed more than 80% of juveniles at all dilutions, while the same level of mortality was obtained with *Acacia* sp., *Calotropis procera*, *P. regia*, *Terminalia arjuna* and *T. bellerica* up to 1:20 dilution.

Callistemon lanceolatus, *Lagerstroemia speciosa* and *L. thorelli* had no adverse effect on ei-

TABLE II - In vitro screening of seed extracts for nematocidal activity against *M. javanica* second stage juveniles (J_2).

Plant species	Per cent J_2 mortality, at 27 ± 1 °C, after 24 hrs			
	Dilutions			Control
	1:10	1:20	1:40	
<i>Acacia eburnea</i>	100	100	91	0
<i>Cassia</i> sp.	100	96	87	0
<i>Sesbania sesban</i>	100	88	71	0
<i>Ponciana regia</i>	100	81	66	0
<i>Acacia</i> sp.	100	70	61	0
<i>Tribulus terrestris</i>	98	93	79	0
<i>Melia azedarach</i>	98	92	83	0
<i>Terminalia arjuna</i>	98	80	64	0
<i>Azadirachta indica</i>	96	59	19	0
<i>Calotropis procera</i>	95	85	62	0
<i>Cassia fistula</i>	93	61	42	0
<i>Terminalia bellerica</i>	92	85	67	0
<i>Cassia occidentalis</i>	90	51	11	0
<i>Chenopodium album</i>	89	52	22	0
<i>Withania somnifera</i>	83	61	20	0
<i>Parthenium hysterophorus</i>	80	60	32	0
<i>Eclipta alba</i>	72	51	28	0
<i>Euphorbia hirta</i>	67	31	1	0
<i>Lantana indica</i>	64	51	21	0
<i>Croton bonplandianum</i>	62	41	12	0
<i>Cannabis sativa</i>	50	31	12	0
<i>Malvastrum coromandelianum</i> ¹	13	6	1	0
<i>Mimusops elengi</i>	12	4	1	0
<i>Nicotiana plumbaginifolia</i>	9	2	0	0
<i>Callistemon lanceolatus</i>	0	0	0	0
<i>Lagerstroemia speciosa</i>	0	0	0	0
<i>Lagerstroemia thorelii</i>	0	0	0	0

¹ With nematostatic effect.

TABLE III - In vitro screening of seed extracts for their inhibitory effects on egg hatch and juvenile (J_2) mortality of *M. incognita*.

Plant species	Per cent egg hatch, at 27 ± 1 °C, after 48 hrs							
	Dilutions						Control	
	1:10		1:20		1:40		Actual egg hatch no.	% J_2 mortality
% Egg ¹ hatch	% J_2 mortality	% Egg hatch	% J_2 mortality	% Egg hatch	% J_2 mortality			
<i>Calotropis procera</i>	1	100	9	85	50	14	87	0
<i>Sesbania sesban</i>	2	100	16	74	80	71	73	0
<i>Cassia occidentalis</i>	5	51	46	49	80	0	41	0
<i>Chenopodium album</i>	10	100	42	58	75	0	28	0
<i>Terminalia bellerica</i>	13	100	48	66	81	0	79	0
<i>Cassia fistula</i>	21	100	55	79	100	36	47	0
<i>Withania somnifera</i>	21	100	59	50	92	0	57	0
<i>Cannabis sativa</i>	23	74	42	53	94	0	105	0
<i>Cassia</i> sp.	25	100	53	50	97	0	100	0
<i>Melia azedarach</i>	28	100	30	74	53	0	56	0
<i>Azadirachta indica</i>	28	100	64	64	88	40	102	0
<i>Terminalia arjuna</i>	31	67	39	52	96	0	91	0
<i>Parthenium hysterophorus</i>	34	82	68	47	88	0	94	0
<i>Tribulus terrestris</i>	36	33	70	29	92	0	82	0
<i>Euphorbia hirta</i>	42	40	57	31	98	0	56	0
<i>Acacia</i> sp.	43	97	76	80	96	2	104	0
<i>Pongamia glabra</i>	48	89	62	55	88	0	94	0
<i>Acacia eburnea</i>	53	100	73	45	96	0	104	0
<i>Croton bonplandianum</i>	53	96	73	36	100	0	60	0
<i>Amaranthus viridis</i>	57	71	59	58	83	0	64	0
<i>Poinciana regia</i>	65	81	90	35	100	12	72	0
<i>Malvastrum coromandelianum</i> ²	76	16	89	6	92	0	74	0
<i>Mimusops elengi</i>	83	8	86	1	98	0	83	0
<i>Parkinsonia aculeata</i>	88	50	90	28	91	0	62	0
<i>Lantana indica</i>	88	6	94	2	96	0	89	0
<i>Lagerstroemia thorelii</i>	91	0	93	0	100	0	100	0
<i>Lagerstroemia speciosa</i>	92	0	98	0	100	0	56	0
<i>Callistemon lanceolatus</i>	97	0	100	0	96	0	39	0

¹ Percentage egg hatch in relation to actual hatch in control.

² With nematostatic effect.

ther *M. incognita* or *M. javanica*, but *Malvastrum coromandelianum* had a nematostatic effect as indicated by the revival of juveniles after being kept in water for a few hours.

Maximum inhibition of egg hatch occurred with the lowest (1:10) dilution of seed extracts. Only *C. procera* (9%), followed by *S. sesban* (16%) and *M. azedarach* (30%) suppressed the

juvenile emergence of *M. incognita* significantly in 1:20 dilution. *S. sesban* was highly toxic to emerged juveniles, causing mortality at all dilutions.

The greatest inhibitory effect on egg hatch of *M. javanica* was shown by *C. procera*, *S. sesban*, *T. arjuna* and *Chenopodium album* at 1:10 dilution. *C. procera* effectively suppressed egg

TABLE IV - In vitro screening of seed extracts for their inhibitory effects on egg hatch and juvenile (J_2) mortality of *M. javanica*.

Plant species	Per cent egg hatch, at $27 \pm 1^\circ\text{C}$, after 48 hrs							
	Dilutions						Control	
	1:10		1:20		1:40		Actual egg hatch no.	% J_2 mortality
	% Egg ¹ hatch	% J_2 mortality	% Egg hatch	% J_2 mortality	% Egg hatch	% J_2 mortality		
<i>Calotropis procera</i>	3	100	24	63	31	32	94	0
<i>Sesbania sesban</i>	3	100	19	85	75	50	56	0
<i>Terminalia arjuna</i>	8	100	48	47	100	0	94	0
<i>Chenopodium album</i>	9	63	28	42	78	0	53	0
<i>Cassia occidentalis</i>	12	100	43	48	97	0	41	0
<i>Terminalia bellerica</i>	12	82	34	12	85	0	101	0
<i>Melia azedarach</i>	13	100	46	37	83	0	73	0
<i>Cassia sp.</i>	14	100	66	46	91	0	104	0
<i>Cannabis sativa</i>	15	81	41	47	88	0	106	0
<i>Withania somnifera</i>	18	100	54	36	95	0	44	0
<i>Euphorbia hirta</i>	22	58	42	49	81	0	77	0
<i>Azadirachta indica</i>	25	100	60	63	92	0	105	0
<i>Cassia fistula</i>	34	100	56	68	85	57	65	0
<i>Parthenium hysterophorous</i>	35	70	71	24	84	0	101	0
<i>Tribulus terrestris</i>	37	39	67	26	95	0	87	0
<i>Acacia eburnea</i>	48	100	68	57	88	33	76	0
<i>Acacia sp.</i>	49	88	69	74	99	0	102	0
<i>Poinciana regia</i>	62	68	75	32	86	16	66	0
<i>Parkinsonia aculeata</i>	63	40	76	32	79	0	69	0
<i>Pongamia glabra</i>	68	49	83	41	98	0	67	0
<i>Croton bonplandianum</i>	69	100	80	50	91	0	36	0
<i>Malvastrum coromandelianum</i> ²	76	14	76	3	96	0	76	0
<i>Mimusops elengi</i>	79	8	87	2	95	0	105	0
<i>Amaranthus viridis</i>	83	57	92	23	93	0	81	0
<i>Lantana indica</i>	86	8	89	1	98	0	75	0
<i>Callistemon lanceolatus</i>	89	0	91	0	95	0	46	0
<i>Lagerstroemia thorelii</i>	94	0	96	0	100	0	82	0
<i>Lagerstroemia speciosa</i>	96	0	96	0	97	0	94	0

¹ Percentage egg hatch in relation to actual hatch in control.

² With nematostatic effect.

hatch at all dilutions (Table IV). *Cassia fistula* and *S. sesban* were most effective against emerged juveniles.

Discussion

Seed extracts of *Acacia eburnea*, *Cassia* sp. and *S. sesban* showed the greatest nematicidal effect at the three concentrations tested against both *M. incognita* and *M. javanica*. Nematicidal specificity of some seeds (*A. indica*, *M. azedarach*, *T. terrestris* and *Eclipta alba*) as indicated by difference in responses of two nematode species needs more trials to be established conclusively.

Inhibitory potential of the extracts does not necessarily correspond with their ability to induce juvenile mortality. Some seed extracts causing high juvenile mortalities did not always inhibit egg hatch effectively, e.g., *A. eburnea*. Besides, the same degree of toxicity was not obtained for mortality of emerging juveniles as compared to when J₂ were exposed directly. The difference could be that continuous hatching of juveniles does not expose all of them to equal or sufficient exposure time. Against *M. incognita*, eleven seeds and against *M. javanica*, twelve seeds allowed less than 30% egg hatch (at 1:10). In general, the extracts were less effective against egg hatch as compared to mortality of emerged juveniles. *C. procera* and *S. sesban* were the most inhibitory for both species. All seeds that were highly effective in suppressing egg hatch, also caused 100% or nearly 100% juvenile mortality, but as the juvenile emergence was low, very few juveniles were actually exposed. Where egg hatch inhibition was not significant, the juvenile mortality rates were important, e.g., *A. eburnea* did not inhibit egg hatch in both species but effectively killed the emerged juveniles, thus indicating the different nature of nematicidal activity. For *M. javanica*, *Croton bonplandianum* and

Acacia sp. also showed similar results. Interestingly, *Cannabis sativa* seeds inhibited egg hatch for both species but were much less effective in causing juvenile mortality.

Callistemon lanceolatus, *L. speciosa* and *L. thorelii* were ineffective in suppressing hatch or in causing mortality. The results clearly indicate the superior potential of seeds of some families, prominently, Leguminosae, Meliaceae and Asclepiadaceae, to inhibit hatch or increase juvenile mortality.

The inhibition of egg hatch and juvenile mortality of *M. incognita* by *M. azedarach* seed extracts was also reported by Lee (1990). High nematotoxic potential of seeds of *S. sesban*, *M. azedarach* and *C. procera* is in conformity with earlier observations in this laboratory, wherein also the seed extracts of *M. azedarach* and *Ricinus communis* L. were found to be more toxic as compared to leaves. Higher potential of seeds of *Leucaena leucocephala* in comparison to other plant parts was also indicated by Jain and Hasan (1984).

In view of the present findings, there is need for extensive screening of seed extracts from many plant species. The immediate application of seed extracts may be as spot, nursery or root-dip treatments, but comprehensive testing is needed for field applications.

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